

## Design, synthesis, and biological evaluation of novel aryl isoxazoles as potential Hsp90 inhibitors

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### Abstract

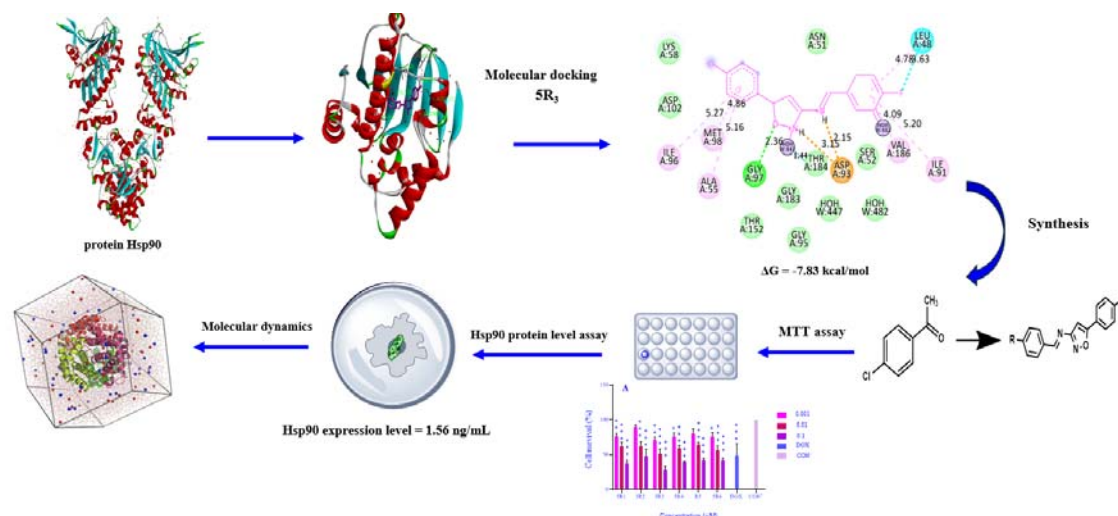
**Background and purpose:** Heat shock protein 90 (Hsp90) is a molecular chaperone critical for the stabilization of numerous oncoproteins, making it a promising target for anticancer drug development. This study aimed to design, synthesize, and evaluate novel 3,5-diarylisoxazole derivatives as potential Hsp90 inhibitors

**Experimental approach:** A series of 3,5-diarylisoxazole compounds (5R<sub>1-6</sub>) was designed using molecular docking to predict binding affinity. Compounds were synthesized *via* cyclization of 4-chloroacetophenone with hydroxylamine hydrochloride, followed by condensation with various aldehydes. Structures were confirmed by melting point, FT-IR, and <sup>1</sup>H-NMR spectroscopy. The MTT assay assessed cytotoxicity, and Hsp90 inhibitory activity was evaluated using an Hsp ELISA kit. The stability of the 5R<sub>3</sub> compound in the Hsp90 active site was investigated using molecular dynamics simulation.

**Findings/Results:** All compounds showed favorable binding energies. Compound 5R<sub>3</sub> showed the highest binding affinity ( $\Delta G = -7.83$  kcal/mol) and the highest cytotoxicity against MCF-7 cells ( $IC_{50} = 0.014$   $\mu$ M) and significantly reduced Hsp90 concentration from 5.54 ng/mL (untreated) to 1.56 ng/mL. Furthermore, molecular dynamics simulation studies confirmed the stability of the Hsp90-5R<sub>3</sub> complex during a 100 ns simulation.

**Conclusion and implications:** Compound 5R<sub>3</sub> with an electronegative substituent (F) on the aromatic ring showed the highest cytotoxic effect, also decreased the concentration of Hsp90 protein more than others.

**Keywords:** Hsp90 inhibitor; Isoxazole; Molecular docking; Molecular dynamics simulation; MTT assay.



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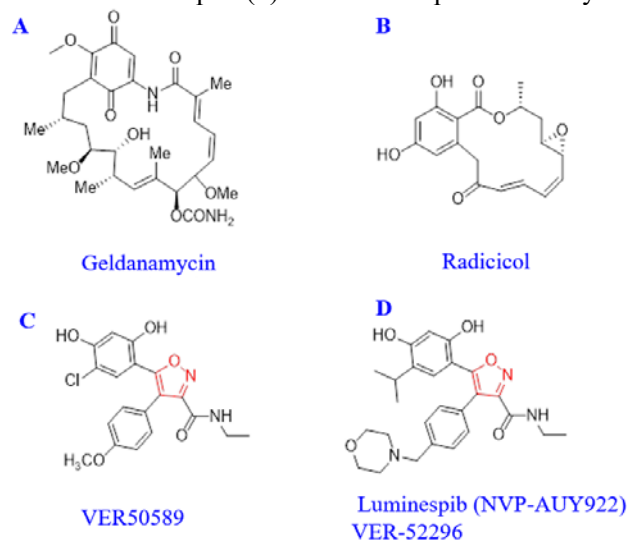
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## INTRODUCTION

Heat shock proteins (Hsps) are highly conserved molecular chaperones involved in protein folding, stabilization, and degradation, particularly under stress conditions. Hsps are classified into several families based on their approximate molecular weights, including Hsp100, Hsp90, Hsp70, Hsp60, and small Hsps, each playing distinct roles in protein folding and cellular stress response (1,2). Hsp90, a key member of this family, plays a crucial role in the folding and activation of numerous client proteins implicated in cancer progression. Inhibition of Hsp90 disrupts the stability of these oncoproteins, making it an attractive target for anticancer therapy (3,4). Hsp90 is a homomeric protein composed of two identical subunits with three domains: an N-terminal ATP-binding domain, a central chaperone domain, and a C-terminal dimerization domain. Most Hsp90 inhibitors target the ATP-binding site in the N-terminal domain. Natural inhibitors such as radicicol and geldanamycin, as well as synthetic compounds like luminespib (NVP-AUY922), have demonstrated potent Hsp90 inhibitory activity (5,6).

Radicicol and geldanamycin (Fig. 1) are introduced as the first natural Hsp90 inhibitors. These compounds work by binding directly to the ATP-binding pocket in the N-terminal domain, effectively competing with ATP and disrupting the associated function of Hsp90 (7).

Besides these natural compounds, numerous novel compounds have been synthesized that act as Hsp90 inhibitors, especially for anticancer therapy (8). Some recent studies highlight heterocyclic molecules such as pyrazole, indole, pyrimidine, triazole, isoxazole, and thioquinazoline that have shown promising anticancer activity (8-12). Luminespib, containing an isoxazole ring, is a high-affinity Hsp90 inhibitor with an  $IC_{50}$  of 21 nM and a  $K_d$  of 1.7 nM (Fig. 1). Its impressive potency has led to its evaluation in phase II of clinical trials (7,13). Most of the previously synthesized isoxazole derivatives bear amide or ketone functionalities at positions 3, 4, or 5 of the isoxazole ring, enabling their conjugation with aryl groups. To enhance their cytotoxic potential, certain compounds feature substitutions exclusively at the 3 and 5 positions, where the substituents are either attached directly to the ring or connected via methylene, ethylene, or cyclic linkers. Galiskan *et al.* reported the synthesis of a series of 3,5-diaryl derivatives incorporating a piperazinyl linker, which exhibited cytotoxic activity against the MCF-7 breast cancer cell line within the concentration range of 1.2-20  $\mu$ M (14). Inspired by the therapeutic potential of isoxazole-based organic molecules (15,16) and our previous works (17,18), we designed and synthesized a novel series of 5-phenylisoxazol-3-amine derivatives to evaluate their cytotoxic and Hsp90 inhibitory activities.



**Fig. 1.** The chemical structure of some natural (A and B) and synthetic Hsp90 inhibitors with an isoxazole ring (C and D) is being investigated in different stages of clinical trial studies.

## MATERIALS AND METHODS

### Compounds and instrumentation

All chemicals used in these experiments were sourced from Sigma-Aldrich or Merck and employed without additional purification. Thin-layer chromatography (TLC) was employed to follow the progress of the reaction and confirm the identity and purity of the substances during the experiment. The melting point of the compounds was measured with an electrothermal melting point instrument model 9200. Infrared spectra were obtained by a JASCO Fourier transform infrared spectroscopy (FT-IR, JASCO, Japan) using KBr discs. Compounds were analyzed *via* proton nuclear magnetic resonance (<sup>1</sup>H-NMR) spectroscopy, conducted with a Bruker Ultra Shield 400 MHz (Bruker, USA) and recorded in CDCl<sub>3</sub> solvents.

### Molecular docking

In this study, experimental docking of 5-phenylisoxazol-3-amine derivatives as Hsp90 inhibitors was performed using AutoDock 4.2 software (19). The protein structure was obtained from the Protein Data Bank (PDB: 5xqd) (20). Its structure was produced by eliminating all water molecules, except for the water molecules critical for protein-ligand interactions, ions, and ligands. Atomic charges were incorporated and computed *via* the Kollman procedure. The file was saved in pdbqt format. A grid box size of 60 × 60 × 60 Å, centered on the protein binding pocket, with a grid spacing of 0.375 Å, was utilized. The structures of the 3D ligands were illustrated using Marvin Sketch v5.7 software and saved in PDB format. The Gasteiger-Marsili method was employed to compute the partial atomic charges of the ligands. This calculation is crucial as partial charges significantly influence molecular binding by dictating macromolecule-ligand interaction (21). The Lamarckian genetic evolution was executed over 100 iterations. Additionally, parameters about water molecules were integrated into the AD4-parameter and AD4-bound files. All runs were categorized according to the lowest binding energy, and a comprehensive analysis was performed to determine the best conformation and ligand orientation within the protein's active site. The docking procedure was analyzed and visualized

utilizing Pymol and Discovery Studio Visualizer version 17.2 (22,23).

### Molecular dynamics simulation

Molecular dynamics (MD) simulations were conducted to investigate the stability of the ligand-Hsp90 complex. Following the biological assessments, compound 5R3 was chosen for MD simulations. The simulations aimed to evaluate the stability of 5R<sub>3</sub> within the Hsp90 active site and to compare its interaction pattern with that of the reference Hsp90 inhibitor AUY-922. MD simulations were performed with GROMACS 2021.5. (24). Protein pKa values were computed with the PROPKA 3.1 web server (25). Protein topology files were prepared using the Amber force field. Ligand topologies were prepared using ACPYPE, which interfaces with antechamber to derive force-field parameters for small molecules (26). Simulations under physiological conditions were performed by solvating the Hsp90-ligand complex in a dodecahedral box containing TIP3P water, followed by the addition of counterions (Na<sup>+</sup> or Cl<sup>-</sup>) to achieve charge neutrality. After energy minimization, a two-step equilibration protocol was applied before production MD. The first step was an NVT equilibration at 323 K for 500 ps, and the second step was an NPT equilibration at 323 K and 1 bar for 500 ps. Following system stabilization, a 100 ns MD simulation was conducted at 323 K. Trajectory analyses were examined for time-dependent properties, including backbone root mean square deviation (RMSD), root mean square fluctuations (RMSF), and radius of gyration (Rg).

### General synthesis procedure of compounds

#### Synthesis of (Z)-3-chloro-3-(4-chlorophenyl)acrylonitrile (2)

Phosphoryl chloride (POCl<sub>3</sub>; 16.6 mmol, 1.6 mL) was added dropwise to 13 mL of dry dimethylformamide (DMF) while keeping the temperature below 25 °C using an ice bath, and it was stirred for 30 min. Then 4-chloroacetophenone (8.3 mmol, 1.2 mL) was added dropwise, while the temperature was maintained at 60 °C. After that, the reaction mixture was stirred at room temperature for 2 h. Then, hydroxylamine hydrochloride (33.32 mmol, 2.1 g) was added dropwise (exothermic reaction), and the reaction was stirred for

another 24 h at room temperature. The progress of the reaction was followed by TLC. After the completion of the reaction, the mixture was poured into crushed ice to obtain a solid product. The resulting product was filtered, washed with cold ethanol, dried, and recrystallized from ethanol to obtain an off-white solid product (76% yield) (27,28).

*Synthesis of (4-chlorophenyl) isoxazol-3-amine (3)*  
(Z)-3-chloro-3-(4-chlorophenyl)

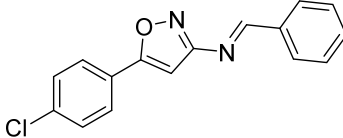
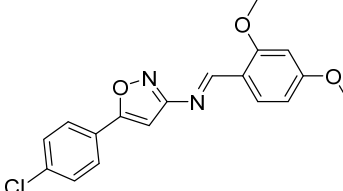
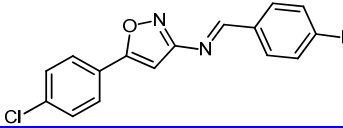
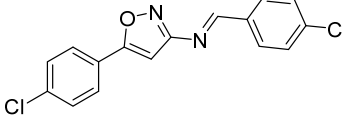
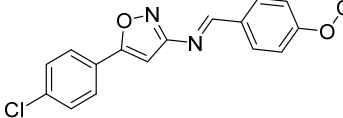
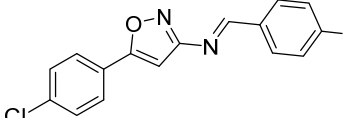
acrylonitrile (2) (10 mmol, 1.2 g) was dissolved in ethanol (10 mL) and stirred for a few min. Sodium hydroxide (96%) and hydroxylamine hydrochloride (15 mmol, 1.5 g) were then added, and stirred to mix while the pH was kept between 7 and 8. The reaction was then heated to 60 °C and stirred for another 24 h. HCl (3 mmol) was added in one portion, and the reaction was heated for 2 h at 80 °C. Then, the reaction was allowed to cool down to room

temperature. The aqueous layer was extracted with EtOAc (2 × 200 mL). The organic layer was dried using MgSO<sub>4</sub> and evaporated to crystallize. Finally, the product was recrystallized from ethanol to give a white powder with a 78% yield (29,30).

*Synthesis of (4-chlorophenyl) isoxazol-3-amine derivatives coupled with various aldehydes (5R<sub>1-6</sub>)*

The final Schiff base products were prepared by mixing equimolar amounts (1 mmol) of the aldehyde derivatives (4) and (4-chlorophenyl) isoxazol-3-amine (3) in absolute ethanol (10 mL) under reflux. After the reaction was completed (checked by TLC), the solvent volume was reduced by evaporation, cooled down to room temperature, and filtered. The obtained solid product was washed several times with cold methanol and recrystallized from a mixture of ethanol and water to give compounds 5R<sub>1-6</sub> as illustrated in Table 1.

**Table 1.** The structure, chemical formula, yield, and melting point of synthesized compounds 5R<sub>1-6</sub>

Compound code	Structure	Chemical formula	Purified compounds yield (%)	Melting point (°C)
5R <sub>1</sub>		C <sub>16</sub> H <sub>11</sub> ClN <sub>2</sub> O	25	195
5R <sub>2</sub>		C <sub>18</sub> H <sub>15</sub> ClN <sub>2</sub> O <sub>3</sub>	25	247
5R <sub>3</sub>		C <sub>16</sub> H <sub>10</sub> ClFN <sub>2</sub> O	55	233
5R <sub>4</sub>		C <sub>16</sub> H <sub>10</sub> Cl <sub>2</sub> N <sub>2</sub> O	45	231
5R <sub>5</sub>		C <sub>17</sub> H <sub>13</sub> ClN <sub>2</sub> O <sub>2</sub>	37	235
5R <sub>6</sub>		C <sub>17</sub> H <sub>13</sub> ClN <sub>2</sub> O	40	224

## Biological evaluations

### Cell lines

Cancerous cells, including HeLa (cervical cancer) and MCF-7 (breast cancer), and a normal cell line, HUVEC, were purchased from the Pasteur Institute (Tehran, Iran). HeLa cells were maintained in Dulbecco's Modified Eagle Medium (DMEM), while HUVEC and MCF-7 cells were maintained in RPMI 1640 medium, at 37 °C in a humidified atmosphere (containing 5% CO<sub>2</sub>). Media were completed with 10% fetal bovine serum (FBS), 100 units/mL penicillin, and 100 µg/mL streptomycin.

### Cytotoxic activity assay

MTT assay was performed to evaluate cell viability and cytotoxic effects of the tested compounds against cancer cells as reported before (31). Briefly, cells were seeded ( $5 \times 10^3$  cells/well) in 96-well plates and incubated for 24 h. After that, the cells were treated with different concentrations of synthesized compounds (0.001, 0.01, and 0.1 µM) and incubated for 48 h. After the exposure period, 20 µL of MTT reagent (5 mg/mL) was added to each well, and incubation continued for an additional 3 h. Subsequently, 150 µL of dimethyl sulfoxide (DMSO) was replaced with media to dissolve the formed formazan crystals, and absorbance was measured at 570 nm using an enzyme-linked immunosorbent assay (ELISA) plate reader (BioTek, USA). Experiments were conducted in triplicate, and cell viability was calculated by comparing treated samples against negative controls. Doxorubicin (2 µg/mL) served as the positive control to validate assay performance.

### Hsp90 protein level assay

The effect of synthesized cytotoxic agents on Hsp90 expression levels was evaluated using the Hsp90 expression level kit (Cusabio, USA), in accordance with the manufacturer's protocol. Standards and samples were added to the designated wells with a concentration of one-fourth of IC<sub>50</sub> values and incubated at 37 °C for 2 h. After incubation, the supernatant was discarded and replaced with 100 µL of biotin-conjugated antibody, followed by a 1-h incubation at 37 °C. The wells were then aspirated and washed three times with 400 µL of the provided wash buffer.

Subsequently, 100 µL of horseradish peroxidase-conjugated avidin (HRP-avidin) was added to each well and incubated for another hour at 37 °C. The washing procedure was repeated as described above. Then, 90 µL of 3,3',5,5'-tetramethylbenzidine (TMB) substrate solution was introduced into the wells and allowed to react for 15-30 min at 37 °C. The reaction was terminated by adding the stop solution, and absorbance was measured at 450 nm using an ELISA plate reader (BioTek, USA).

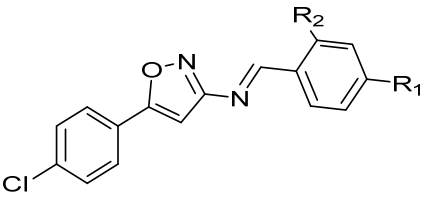
### Statistical analysis

IC<sub>50</sub> values were calculated by linear regression and expressed as mean ± SD. All linear regressions in this paper were analyzed using GraphPad Prism 6 software (32). Statistical analysis was performed using one-way analysis of variance (ANOVA) followed by Tukey's post hoc test to determine the significant differences among the groups. \* $P \leq 0.05$  values were considered statistically significant.

## RESULTS

### Docking studies

Molecular docking analysis was performed to investigate the binding interactions between the drugs and the Hsp90 binding site. The results, including binding free energies ( $\Delta G$ ), electrostatic interactions, hydrogen binding, and hydrophobic interactions, are summarized in Table 2. To validate the docking protocol, the NVP-AUY922 compound, as a reference and crystallographic ligand were redocked. Co-crystal ligand occupies the Hsp90 active site in a manner that is surrounded by essential amino acids such as Asp93, Gly97, Thr184, Ala55, Val186, Met98, Ile91, and Ile96, along with structural water molecules. A crystallographic ligand with binding energies ( $\Delta G$ : -7.09) and a hydrogen bond is formed with the oxygen of the isoxazole ring with amino acid Gly97 at a distance of 2.86 Å and amino acid Asp93 at a distance of 1.69 Å. In compound NVP-AUY922, hydrogen bonds are formed by the oxygen group of the isoxazole ring with amino acid Thr184 at a distance of 2.04 Å and amino acid Asp93 at a distance of 1.80 Å, and by water molecule 447 at a distance of 2.70 Å and water molecule 482 at a distance of 2.55 Å.

**Table 2.** Binding energy ( $\Delta G$ ), hydrophobic and hydrophilic interactions of synthesized compounds and NVP-AUY922 in the active site of Hsp90 protein.


Compounds	R <sub>1</sub>	R <sub>2</sub>	( $\Delta G$ ) (kcal/mol)	H-Bond amino acids (distance Å <sup>0</sup> )	Electrostatic amino acids	Hydrophobic Amino Acids
Luminespib (NVP-AUY922)	-	-	-12.42	Gly97 (2.27), Thr184 (2.09), Asp93 (1.80)	Met98	Ala55, Lys58, Ile96, Val186
Co-crystal ligand	-	-	-7.09	Gly97 (2.86), Asp93 (1.69)	Met98	Ala55, Lys58, Ile96
5R <sub>1</sub>	H	H	-6.78	Leu107 (2.14), Thr184 (1.59)	Asp102, Met98	Leu48, Val186, Ile96
5R <sub>2</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	6.47-	Gly97 (2.26), Lys58 (2.51)	Asp102, Met98	Ala55
5R <sub>3</sub>	F	H	-7.83	Gly97 (2.36), Thr184 (1.44)	Asp93	Ala55, Val186, Met98, Ile91-96
5R <sub>4</sub>	Cl	H	-6.78	Leu107 (2.09)	Asp93, Met98	Ile96, Val186, Leu48
5R <sub>5</sub>	OCH <sub>3</sub>	H	-6.43	Leu107 (2.12)	Met98	Ala55
5R <sub>6</sub>	CH <sub>3</sub>	H	-6.65	Leu107 (2.15)	Met98	Val186, Ile96

Hydrophobic interactions of the ligand with Ala55, Lys58, Ile96, Val186, and the formation of electrostatic interactions with Met98 are shown in Fig. 2. The RMSD between the experimental and predicted states was calculated. The RMSD value obtained was 1.4 Å, indicating good accuracy and reliability of the docking method.

### Synthesis of compounds

The target compounds were synthesized through a three-step synthetic route as illustrated in the synthetic scheme (Fig. 3).

In step 1, the initial ketone (compound 1) was subjected to the Vilsmeier-Hack reaction using DMF and POCl<sub>3</sub> in the presence of hydroxylamine hydrochloride to produce intermediate compound 2, containing a chlorinated aromatic moiety and a cyano substituent (33). Step 2 involved the conversion of compound 2 to the isoxazole derivative 3 via reaction with sodium hydroxide and ethanol in the presence of hydroxylamine hydrochloride, which resulted in ring closure and formation of the amino-isoxazole scaffold. Finally, in step 3, compound 3 was subjected to condensation with various substituted benzaldehydes

(compound 4) in ethanol under acidic conditions to provide the final Schiff base derivatives 5R<sub>1-6</sub>, as shown in the Fig. 3. This multi-step synthesis efficiently provided the desired compounds with the designed substitution pattern. The lower yields obtained for some final derivatives can be explained by side-product formation and purification difficulties.

#### (E)-N-(5-(4-chlorophenyl)isoxazol-3-yl)-1-phenylmethanimine (5R<sub>1</sub>)

C<sub>16</sub>H<sub>11</sub>ClN<sub>2</sub>O; molecular weight (MW): 282.73 Da; melting point (m.p.): 195 °C; yield: 25%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1622 (N=CH); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.59 (s, 1H, N=CH), 7.63 (m, 4H, Ar), 7.47 (m, 5H, Ar), 6.04 (s, 1H, CH).

#### (E)-N-(5-(4-chlorophenyl)isoxazol-3-yl)-1-(2,4-dimethoxyphenyl)methanimine (5R<sub>2</sub>)

C<sub>18</sub>H<sub>15</sub>ClN<sub>2</sub>O<sub>3</sub>; MW: 342.78 Da; m.p.: 247 °C; yield: 25%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1622 (N=CH); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.75 (d, *J* = 2.2 Hz, 1H, N=CH), 7.68 (m, 2H, Ar), 7.46 (m, 3H, Ar), 7.40 (m, 2H, Ar) 6.96 (d, *J* = 8.4 Hz, 1H, CH), 3.99 (d, *J* = 10.7 Hz, 6H, OCH<sub>3</sub>).



*(E)-1-(4-chlorophenyl)-N-(5-(4-chlorophenyl)isoxazol-3-yl)methanimine (5R3)*

C<sub>16</sub>H<sub>10</sub>Cl<sub>2</sub>N<sub>2</sub>O; MW: 317.17 Da; m.p.: 231 °C; yield: 55%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1622 (N=CH); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.83 (s, 1H, N=CH), 7.93 (m, 2H, Ar), 7.78 (dd, *J* = 9.0, 2.6 Hz, 2H, Ar), 7.51 (d, *J* = 7.5 Hz, 4H, Ar), 6.64 (d, *J* = 2.9 Hz, 1H, CH).

*(E)-N-(5-(4-chlorophenyl)isoxazol-3-yl)-1-(4-fluorophenyl)methanimine (5R4)*

C<sub>16</sub>H<sub>10</sub>ClF<sub>2</sub>N<sub>2</sub>O; MW: 300.72 Da; m.p.: 233 °C; yield: 45%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1607 (N=C); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.83 (s, 1H, N=CH), 7.94 (dd, *J* = 8.6, 2.3 Hz, 2H, Ar), 7.78 (dd, *J* = 9.0, 2.6 Hz, 2H, Ar), 7.51 (m, 4H, Ar), 6.64 (d, *J* = 2.8 Hz, 1H, CH).

*(E)-N-(5-(4-chlorophenyl)isoxazol-3-yl)-1-(4-methoxyphenyl)methanimine (5R5)*

C<sub>17</sub>H<sub>13</sub>ClN<sub>2</sub>O<sub>2</sub>; MW: 312.75 Da; m.p.: 235 °C; yield: 37%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1607 (N=C); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.81 (s, 1H,

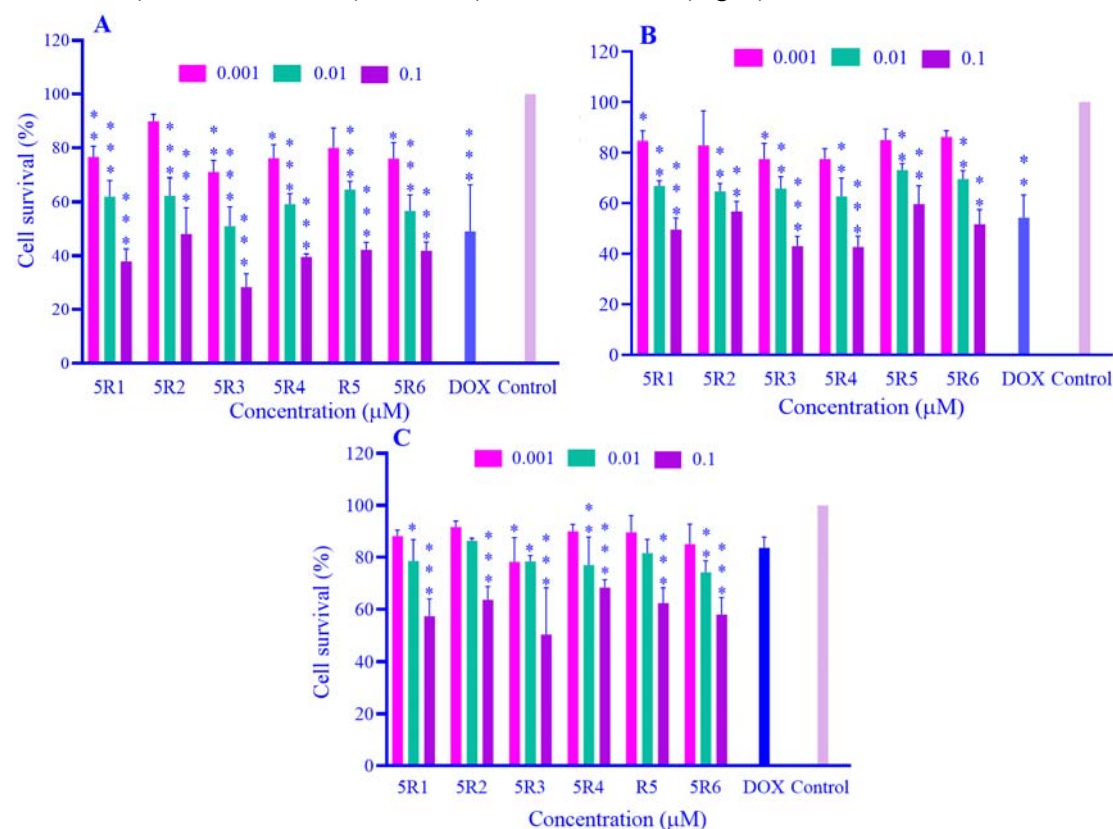
N=CH), 7.84 (dd, *J* = 8.8, 3.5 Hz, 2H, Ar), 7.70 (m, 2H, Ar), 7.12 (m, 2H, Ar), 7.04 (m, 2H, Ar), 6.18 (s, 1H, CH), 3.81 (s, 1H, OCH<sub>3</sub>).

*(E)-N-(5-(4-chlorophenyl)isoxazol-3-yl)-1-(p-tolyl)methanimine (5R6)*

C<sub>17</sub>H<sub>13</sub>ClN<sub>2</sub>O; MW: 296.75 Da; m.p.: 224 °C; yield: 40%; IR (KBr):  $\nu$  (cm<sup>-1</sup>) 1607 (N=C); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.24 (m, 1H, N=CH), 8.09 (m, 2H, Ar), 7.69 (m, 2H, Ar), 7.56 (t, *J* = 7.9 Hz, 2H, Ar), 7.36 (m, 2H, Ar), 6.64 (s, 1H, CH), 2.78 (s, 3H, CH<sub>3</sub>).

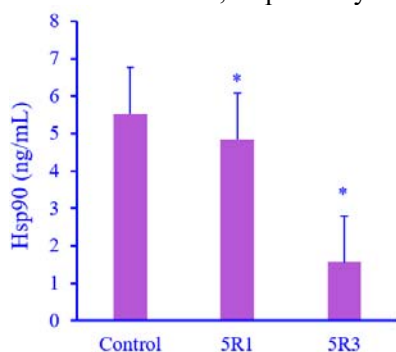
**Biological assays**

The biological activities of the synthesized compounds were assessed through *in vitro* assays, including cytotoxicity evaluation and ELISA-based quantification of Hsp90 expression levels. The antiproliferative effects of the six synthesized compounds were evaluated using the MTT assay against MCF-7, HeLa, and HUVEC cell lines (Fig. 4).

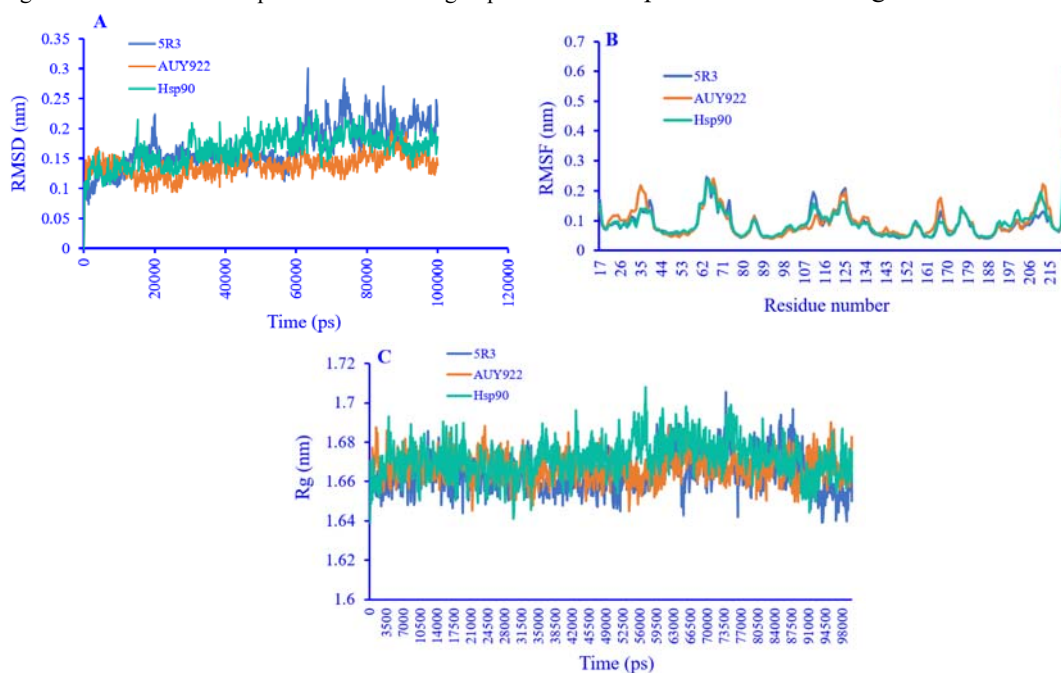


**Fig. 4.** Cytotoxicity assessment of synthesized isoxazole derivatives against (A) MCF-7, (B) HeLa, and (C) HUVEC cell lines using the MTT assay. Data are expressed as mean  $\pm$  SD, *n* = 3. \**P*  $\leq$  0.05, \*\**P*  $\leq$  0.01, \*\*\**P*  $\leq$  0.001 indicate significant differences compared to the control group. DOX, Doxorubicin.

As shown in Fig. 4, among the six compounds, 5R3 showed the highest antiproliferative activity with IC<sub>50</sub> of  $0.014 \pm 0.002 \mu\text{M}$ ,  $0.050 \pm 0.010 \mu\text{M}$ , and  $0.150 \pm 0.030 \mu\text{M}$  against MCF-7, HeLa, and HUVEC, respectively. We used an ELISA kit to detect the intracellular Hsp90 protein level. For this purpose, the compounds with the highest cytotoxicity (5R<sub>3</sub> and 5R<sub>4</sub> at a concentration of  $\times 0.25 \text{ IC}_{50}$ ) were used and the results are shown in Fig. 5. As seen in this figure, the level of expression of Hsp90 proteins was reduced from  $5.5 \pm 1.10 \text{ ng/mL}$  in untreated cells (negative control) to  $1.5 \pm 0.15$  and  $4.8 \pm 0.70 \text{ ng/mL}$  by compounds 5R<sub>3</sub> and 5R<sub>1</sub>, respectively.



**Fig. 5.** Inhibitory effects of compound 5R<sub>1</sub> and 5R<sub>3</sub> on Hsp90 protein levels in MCF-7. Un-treated cells were used as a negative control. Data represent the mean  $\pm$  SD of three independent experiments ( $n = 3$ ). \* $P \leq 0.05$  indicates significant differences compared to the control group.



**Fig. 6.** (A) RMSD plot; (B) RMSF plot; and (C) Rg plot of the Hsp90 protein backbone in complex with compounds 5R<sub>3</sub>, AUY-922, and Hsp90 protein. RMSD, Root mean square deviation; RMSF, root mean square fluctuations; Rg, radius of gyration.

### MD simulation

A 100 ns MD simulation was conducted. To evaluate backbone stability of the Hsp90-ligand complexes, the RMSD was computed for the ligand 5R<sub>3</sub>, the Hsp90 protein, and the reference inhibitor AUY-922. As shown in Fig. 6A, the RMSD trajectories for the 5R<sub>3</sub>-Hsp90 complex, the Hsp90 protein, and the Hsp90 inhibitor were nearly superimposable in the initial 50 ns, indicating similar early-stage conformational stability. RMSD analysis shows that AUY922 bound to Hsp90 exhibits greater stability than both apo Hsp90 and Hsp90 in complex with 5R<sub>3</sub> for the remainder of the simulation. Notably, 5R<sub>3</sub> displays fluctuations under 0.3 nm, reflecting its stability in the Hsp90 active site. Figure 6B presents the RMSF of the Hsp90 protein, where lower values correspond to greater residue stability. The active-site residues Asn51, Met98, Ala55, Asp93, Leu107, Phe138, Asn106, and Thr184 exhibited stable behavior across the three independent simulations, indicating stability of the backbone in the Hsp90 active site. Figure 6C presents the Rg for the two Hsp90-ligand complexes and the apo protein. The Rg trajectories were highly similar across systems, with conserved coherence over the entire simulation, suggesting compactness of the Hsp90 backbone during MD simulation.

## DISCUSSION

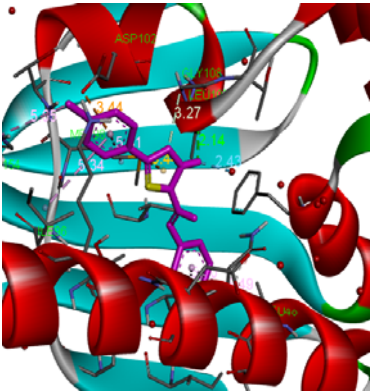
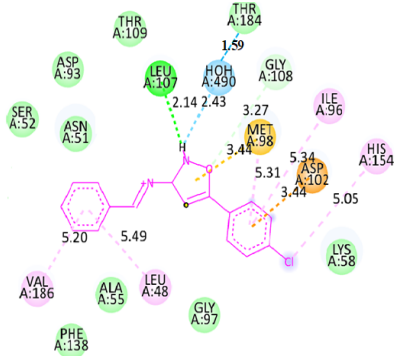
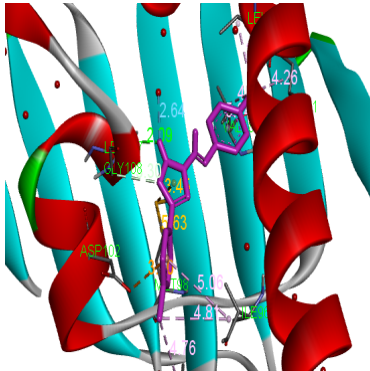
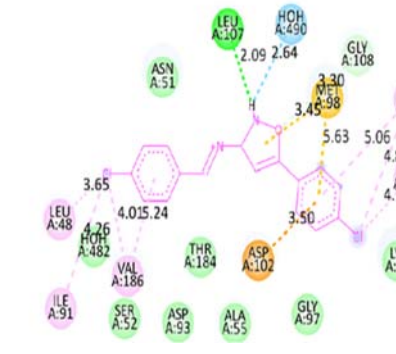
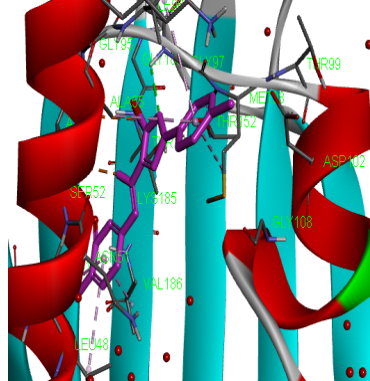
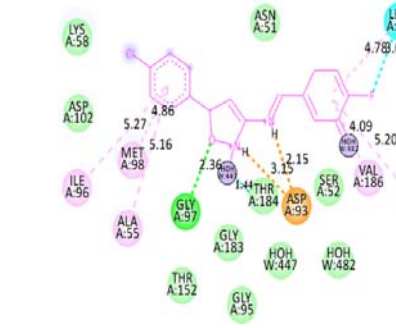
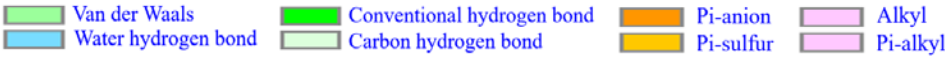
Most previously synthesized isoxazole derivatives feature the isoxazole ring linked to aryl groups directly or *via* amide bonds to an aliphatic group; however, ethylenic and imine linkages have also been reported, occasionally enhancing biological activity. For example, converting the two carbonyl groups in curcumin into an isoxazole ring significantly improved its cytotoxicity against MCF-7 cells, reducing IC<sub>50</sub> from 21.89 to 3.97  $\mu$ M (34). Balaji *et al.* conjugated Hispolon with an isoxazole ring, resulting in enhanced anti-tuberculosis activity (MIC = 1.6  $\mu$ g/mL) (35). Yin and colleagues linked isoxazole to chalcone and observed moderate activity against melanoma cells (IC<sub>50</sub> = 135  $\mu$ M) (36). Arylation of isoxazole *via* imine bonds at positions 3 and 4 produced compounds more potent than erlotinib against A549 cells, with IC<sub>50</sub> improving from 25 to 17  $\mu$ M (37). Additionally, conjugation with N-aryl pyrazole yielded a derivative exhibiting strong cytotoxicity (IC<sub>50</sub> = 0.31  $\mu$ M) toward MCF-7 cells (38). Furthermore, our compounds are 5-aryl substituted isoxazoles that contain fluorine and chlorine, electron-rich substituents, that create stronger electrostatic and hydrophobic interactions with additional residues such as Asn51, Asp58, Met98, and Val150 in the Hsp90 pocket. Mechanistically, unlike previous inhibitors, our fluorinated analogs significantly reduce Hsp90 expression in cancer cells (down to 1.56 ng/mL) while maintaining lower cytotoxicity in normal cells, indicating a distinct modulation profile of the chaperone cycle rather than a complete ATP-competitive inhibition (38).

Imines, especially those attached to aromatic rings, have shown significant potential as anticancer agents through various mechanisms such as induction of apoptosis, production of reactive oxygen species (ROS), and disruption of vital cellular functions (39,40). They have also shown the ability to suppress the growth of MCF-7 cells and are considered promising anticancer candidates (41). The imine group, with its C=N double bond, is highly polar, allowing interaction with key amino acid residues such as Asp93, Thr184, and Lys5 (42). Imines can also act as intermediate rings

between aryl or heterocyclic groups and are relatively stable in aqueous and cellular environments, making them suitable for the design of selective and cell-penetrating inhibitors (40). Building upon previous findings and our molecular docking studies of the designed compounds, and considering the novelty of the approach, this study focused on synthesizing isoxazole derivatives functionalized with aryl groups *via* imine bonds at positions 3 and 5, followed by evaluation of their cytotoxicity and effects on Hsp90 protein expression.

The interaction of the novel isoxazole derivatives with the Hsp90 protein binding site was investigated by docking studies, the dominant method for predicting their function. As shown in Table 2, all synthesized compounds occupy the Hsp90 active site in a manner that is surrounded by essential amino acids such as Asp93, Gly97, Thr184, Ala55, Val186, Met98, Ile91, and Ile96, along with structural water molecules. Among the designed derivatives in compound 5R<sub>1</sub>, hydrogen bonds are formed directly by the NH group of the isoxazole ring with amino acid Leu107 at a distance of 2.14 Å and indirectly by crystallographic water with Thr184 at a distance of 1.59 Å. Hydrophobic interactions of the ligand with Leu48, Val186, and Ile96 and the formation of electrostatic interactions of the isoxazole ring oxygen with Met98 are shown. Compound 5R<sub>4</sub> forms hydrogen bonds *via* the NH group of the isoxazole ring with the amino acid residue Leu107 at a distance of 2.09 Å and with a crystallographic water molecule at a distance of 2.64 Å. This ligand makes hydrophobic interactions with Val186, Leu48, and Ile96 and electrostatic interactions with the amino acid Met98. Compound 5R<sub>3</sub> forms hydrogen bonds directly through the oxygen of the isoxazole ring to amino acid Gly97 at a distance of 2.36 Å and indirectly *via* a crystallographic water to Thr184 at a distance of 1.44 Å. This ligand makes hydrophobic interactions with Ala55, Val186, Met98, Ile91, and Ile96, and the N group makes electrostatic interactions with the amino acid Asp93, which are shown in Table 3. Both the optimized 5R<sub>3</sub> structure ( $\Delta G = -7.83$ ) and the co-crystal structure ( $\Delta G = -7.09$ ) showed acceptable binding energies.

**Table 3.** The illustration of the binding mode of compounds 5R<sub>1</sub>, 5R<sub>3</sub> and 5R<sub>4</sub> (3D and 2D structures) in the active site of Hsp90 protein, derived from AutoDock4.

Compound No.	3D structure	2D structure
5R <sub>1</sub>		
5R <sub>4</sub>		
5R <sub>3</sub>		
<b>Interactions</b>		

The key interactions underlying this stability included hydrogen bonding between the oxygen atom of the isoxazole ring and the amino acid Gly97 in both complexes. In addition, hydrophobic interactions were present with the essential amino acids of the active site, particularly Ala55, Val186, and Ile91-96. The binding  $\Delta G$  of the proposed derivatives was as

follows: NVP-AUY922 > 5R<sub>3</sub> > co-crystal ligand > 5R<sub>1</sub> and 5R<sub>4</sub> > 5R<sub>6</sub> > 5R<sub>2</sub> > 5R<sub>5</sub>.

In this study, six isoxazole derivatives (compounds 5R<sub>1-6</sub>) were synthesized following the procedure outlined in Fig. 3. These imine-functionalized isoxazole derivatives were created by condensing 4-chlorophenylisoxazol-3-amine with various aldehydes.

The presence of the imine group (N=CH) was confirmed through IR spectra, which revealed characteristic absorption bands in the range of 1607-1622  $\text{cm}^{-1}$ , indicating the C=N stretching vibration.

$^1\text{H-NMR}$  spectra for both intermediate and final products were recorded in  $\text{CDCl}_3$ , with proton signal assignments detailed in the experimental section. The aromatic and aliphatic protons of the various derivatives appeared within their expected chemical shift ranges. Notably, the imine proton peaks (N=CH) for the different derivatives resonated between 7.25 and 9.81 ppm, and the aromatic proton peaks of the derivatives were observed between 7.04 and 8.09 ppm.

In this study, the cytotoxic effects of the synthesized isoxazole derivatives were evaluated using the MTT assay against two cancer cell lines (MCF-7 and HeLa), along with the normal human umbilical vein endothelial cells, HUVEC. The  $\text{IC}_{50}$  values were calculated to quantify the potency of each compound (43). Generally, most tested compounds demonstrated significant cytotoxic activity against cancer cell lines at concentrations  $\geq 0.01 \mu\text{M}$ .

As depicted in Fig. 4A, compounds 5R<sub>3</sub>, 5R<sub>1</sub>, and 5R<sub>4</sub> exhibited pronounced cytotoxicity against MCF-7 cells with  $\text{IC}_{50}$  values of  $0.014 \pm 0.002$ ,  $0.029 \pm 0.080$ , and  $0.030 \pm 0.005 \mu\text{M}$ , respectively. Moderate activity was observed for 5R<sub>6</sub>, 5R<sub>5</sub>, and 5R<sub>2</sub>, with  $\text{IC}_{50}$  values of 0.031, 0.045, and 0.063  $\mu\text{M}$ , respectively. Against HeLa cells (Fig. 4), all derivatives (except for 5R<sub>2</sub> and 5R<sub>5</sub>) at concentrations  $> 0.1 \mu\text{M}$  showed cytotoxic effects; however, compounds 5R<sub>3</sub> and 5R<sub>1</sub> showed  $\text{IC}_{50}$  values of  $0.050 \pm 0.01$  and  $0.080 \pm 0.06 \mu\text{M}$ , respectively. Similarly, compounds 5R<sub>2</sub>, 5R<sub>5</sub>, and 5R<sub>6</sub> displayed the lowest cytotoxicity against HeLa cells, with  $\text{IC}_{50}$  values exceeding 0.1  $\mu\text{M}$ . Here, our findings are consistent with previous studies investigating the cytotoxic effects of pyrazole- and isoxazole-based Hsp90 inhibitors such as VER-49009 and VER-50589 (44). In a study by Sharp *et al.* (45), the pyrazole derivative VER-49009 inhibited the growth of the MCF-7 cell line at an effective concentration ( $\text{GI}_{50} = 1360 \text{ nM}$ ), whereas the isoxazole analogue VER-50589 suppressed the same cell line at a much lower concentration ( $\text{GI}_{50} = 197 \text{ nM}$ ), which is

closer to the potency of compound 5R<sub>3</sub> observed in our study ( $\text{IC}_{50} = 14 \text{ nM}$ ). In this regard, Sun *et al.* reported that Luminespib, an isoxazole-containing compound, exhibited cytotoxic activity against MCF-7 and HeLa cells by 50% at concentrations of 6.4 nM and 7.7 nM, respectively (46). In agreement with their findings, our results indicated that the synthesized compounds exhibited significantly higher cytotoxicity toward MCF-7 cells compared to HeLa cells (Fig. 4A and B).

The compounds synthesized in our study contained an isoxazole ring linked to an imine moiety, none of which exhibited cytotoxic effects against the normal HUVEC cell line up to a concentration of 0.1  $\mu\text{M}$ . (Fig. 4C). In contrast, Sharp and colleagues evaluated the compound VER-50589, possessing an isoxazole ring without an imine group, against the same cell line and reported significant cytotoxicity ( $\text{GI}_{50} = 19 \text{ nM}$ ) (45). These findings suggest that isoxazole derivatives conjugated with an imine group may exhibit enhanced selectivity towards cancer cell lines over normal cells.

The cytotoxicity of all compounds tested in this study against the examined cancer cell lines was concentration-dependent, although compounds 5R<sub>3</sub> and 5R<sub>1</sub> demonstrated notably higher efficacy. Based on these findings, these two compounds were identified as promising isoxazole derivatives warranting further investigation.

The intracellular levels of Hsp90 protein were quantitatively measured using a commercially available ELISA kit to evaluate the effect of the synthesized compounds on Hsp90 expression (18). Cytotoxic compounds 5R<sub>1</sub> and 5R<sub>3</sub> were selected for treatment, and their impact on Hsp90 protein concentration in cancer cell lysates was assessed relative to untreated controls. The ELISA results demonstrated a significant reduction in Hsp90 levels in cells treated with 5R<sub>1</sub> and 5R<sub>3</sub>, exhibiting concentrations of  $4.85 \pm 0.70$  and  $1.56 \pm 0.15 \text{ ng/mL}$ , respectively, compared to  $5.54 \pm 1.10 \text{ ng/mL}$  in control cells (Fig. 5). Notably, compound 5R<sub>3</sub> emerged as the most potent Hsp90 inhibitor among the synthesized derivatives (Fig. 5), displaying significant activity at a concentration of  $0.25 \times \text{IC}_{50}$  in MCF-7 cells which was in line with the results

of previous studies (47,48). MD simulations across 100 ns showed that 5R<sub>3</sub> remains firmly positioned in the ATPase active-site pocket, indicating robust binding stability. Supported by its cytotoxic activity and Hsp90 inhibition, the imine-isoxazole derivative 5R<sub>3</sub> emerges as a promising lead compound for the development of novel anticancer therapeutics targeting Hsp90.

## CONCLUSION

In this study, a novel series of 5-aryl isoxazole derivatives containing an imine moiety was rationally designed and synthesized as potential Hsp90 inhibitors. Molecular docking studies demonstrated that these derivatives effectively bind to the Hsp90 active site through a combination of hydrogen bonds, electrostatic, and hydrophobic interactions, underscoring their potential as targeted anticancer agents. Structural elucidation was confirmed *via* IR and <sup>1</sup>H-NMR spectroscopy. The cytotoxicity evaluation revealed that the most potent compound 5R<sub>3</sub>, containing an F pharmacophore moiety, exhibited a strong affinity for Hsp90, lowering its concentration to approximately 1.5 ng/mL and inhibiting MCF-7 cell proliferation with an IC<sub>50</sub> of 0.014 ± 0.002 μM. Additional MD analyses confirmed that these compounds maintained stable occupancy within the Hsp90 ATPase active site throughout the simulation. These promising *in-vitro* outcomes position these isoxazole-imine derivatives as viable leads for optimization in the development of selective Hsp90 inhibitors. Nevertheless, to progress toward clinical relevance, further optimization, comprehensive *in vivo* assessment, and mechanistic investigations are warranted to enhance efficacy, selectivity, and pharmacokinetic profiles.

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## Conflicts of interest statement

All authors declared no conflict of interest in this study.

## Authors' contribution

H. Sadeghi Aliabadi designed and supervised the project. M. Ardestani performed the experiments, analyzed, and interpreted the data. F. Keshavarzipour assisted with spectral interpretation and manuscript writing. M. Abbasi contributed to the molecular docking study. A. Zarghi helped synthesize the compounds. M. Aghaei advised on the biological studies, and M. Ghiaci was the advisor for compound synthesis. All authors have read and approved the finalized article. Each author has fulfilled the authorship criteria and affirmed that this article represents honest and original work.

## AI declaration

In this article, artificial intelligence tools were used to rewrite and refine certain sentences.

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