

Review Article

Exploring the potential of anticancer peptides as therapeutic agents for cancer treatment

Reza Ghavimi^{1,2,3}, Samira Mahmoudi⁴, Mohsen Mohammadi⁵, Elahe Khodamoradi⁶, and Ali Jahanian-Najafabadi^{6,7,*}

¹Division of Biotechnology and Molecular Medicine and Department of Pathobiological Sciences, School of Veterinary Medicine, Baton Rouge, LA, United States.

²CinnaGen Medical Biotechnology Research Center, Alborz University of Medical Sciences, Karaj, I.R. Iran.

³CinnaGen Research and Production Co, Alborz, I.R. Iran.

⁴Department of Biochemistry and Molecular Biology, LSU Health-Shreveport, Shreveport, Louisiana 71104, USA. ⁵The Wistar Institute, 3601 Spruce Street, Philadelphia, PA 19104, USA.

⁶Department of Pharmaceutical Biotechnology, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, I.R. Iran.

⁷Bioinformatics Research Center, Faculty of Pharmacy, Isfahan University of Medical Sciences, Isfahan, I.R. Iran.

Abstract

Despite great advances in cancer identification and treatment, malignancies remain the primary cause of high morbidity and mortality worldwide. The drawbacks of conventional chemotherapy, such as severe toxicity, lack of specificity related to actively dividing cells, and resistance, can warrant the urgent need to develop an alternative approach to treat this disease. To overcome the drawbacks, researchers are attempting to deliver drugs to the site of action (targeted delivery) or to identify drugs that specifically target tumor cells. In this regard, highly cationic and amphipathic antimicrobial peptides are attracting the attention of researchers due to their potent anticancer activity, low cost of manufacture, and, most critically, tumor-targeting activity. A growing number of documents have shown that some of the mentioned peptides exhibited a broad spectrum of cytotoxic activity against cancer cells but not normal mammalian cells entitled as anticancer peptides. Due to their solubility, low toxicity, strong tumor penetration, high selectivity, and ability to be used alone or in conjunction with other conventional medications, anticancer peptides have the potential to become very successful cancer treatments in the future. This review provided an overview of the studies concerning anticancer peptide classification, modes of action, and selectivity, and also summarized some of the anticancer peptides developed for targeting different types of malignancies. The role of in silico methods or artificial intelligence in the design and discovery of anticancer peptides was briefly explained. Additionally, the current review addressed challenges in utilizing anticancer peptides for cancer therapy and highlighted peptides currently undergoing clinical trials.

Keywords: Anticancer peptides; Antimicrobial peptides; Cancer; Cancer therapy; Targeted therapy.

INTRODUCTION

Cancer is a general term used to describe more than 100 distinct, mostly malignant diseases affecting many different tissues and cell types, characterized by the rapid growth of abnormal cells that results from the accumulation few inherited of a or environmentally-induced genetic mutations and epigenetic changes. To become cancerous, a cell must acquire 6 unique behaviors, including I. the ability to produce its growth factors; II. insensitivity to growth-inhibitory signals; III. resistance to cellular suicide mechanisms resulting in apoptosis; IV. the capacity for limitless replication; V. the ability to stimulate new blood vessel development (neovascularization or angiogenesis); and VI. the capacity to invade other organs and tissues, a process known as metastasis.



^{*}Corresponding author: A. Jahanian-Najafabadi Tel: +98-3137927056, Fax: +98-3136680011 Email: Jahanian@pharm.mui.ac.ir

Cancers including breast, lung, liver, prostate, and colorectal are the most commonly diagnosed forms of this disease (1-4). At the beginning of the 21st century, cancer remains one of the leading causes of mortality and morbidity worldwide and continues to take a toll on global public health systems. According to recent reports, it was estimated that 19.3 million new cancer cases and almost 10 million cancer deaths occurred in 2020 worldwide. The illness is the second most common reason for death throughout the world, and per official projections, it is predicted that there will be approximately 26 million new cancer cases and 17 million cancer-related deaths annually by 2030 (2,5). There are some curative therapies available to fight cancer, such as chemotherapy, surgery, and radiotherapy, which play an important role in increasing the life expectancy of cancer patients. Although localized/solid tumors can often be successfully treated by radiation therapy or surgery, chemotherapy is still the principal strategy applied to treat advanced or metastatic cancer. In addition, other therapeutic arsenals. including DNA-alkylating agents, natural products, antimetabolites, hormone agonists/antagonists, and specific inhibitors, such as kinase inhibitors, monoclonal antibodies, or small organic molecules, are available and are used for cancer therapy (6,7).

Current conventional chemotherapeutics, unfortunately, are not specialized for malignant cells and because such therapeutics do not present selective mechanisms to detect normal/abnormal cellular dividing rates, they have severe side effects such as alopecia, vomiting, rashes, and, in some cases, myelosuppression on other cells and organs throughout the body. Little or no tumor specificity, poor tumor penetration, cancer cell heterogeneity, insufficient drug accumulation tumors abound with into the the development of multidrug resistance (MDR) conferred by many factors, and finally, undesirable side effects are all crucial matters that contribute to the lack of effectiveness and eventually therapeutic failure observed in conventional cancer therapeutics. Studies performed on using current remedies have also shown a potential to create secondary malignancies as well as high chances of reoccurrence in many cases. Furthermore, owing to different reasons, a considerable number of cancer patients did not respond to these therapeutics efficiently (8,9). Considering that the number of individuals suffering from cancer-related disorders is growing by the day and since conventional therapies typically have a troubling number of deficiencies and drawbacks, it is critical to establish a new therapeutic strategy. Targeted therapeutic tactics may be useful in this case. Targeted therapy is one of the major modalities in cancer pharmacotherapy, which aims to selectively kill cancer cells and restrict side effects by enhancing the efficacy and specificity of medications. In this scenario, developing a new class of anti-cancer drugs with greater selectivity and specificity against the different types of tumors is highly desirable. Therefore, the efforts of academia and industry are directed toward the prospection of drug-candidate molecules (10,11).

Biopharmaceuticals therapeutic like peptides and proteins, fusion proteins, monoclonal antibodies, and antibody-drug conjugates are the key components of the targeted therapy approaches. Among the mentioned biopharmaceuticals, small bioactive molecules named antimicrobial peptides (AMPs) have drawn the attention of researchers in recent times. AMPs are short peptides and components of the innate immune system that play a vital role in the innate immune system with a wide spectrum of activity against pathogens, including bacteria, viruses, fungi, and parasites. The results of studies from the last decades revealed that some of the AMPs (particularly AMPs with net positive charge) have cytotoxic activity against cancer cells, known as anticancer peptides (ACPs). The electrostatic interaction between negatively charged phosphatidylserine on the surface of cancer cells and positively charged AMPs/ACPs is thought to play a fundamental role in the selectivity of the peptides toward tumors (Fig. 1). As an alternative chemotherapeutic agent, AMPs/ACPs display several extraordinary properties such as broad spectrum activity, high specificity, rapid mode of action, efficient tumor penetration due to

small size, good solubility, low toxicity, ability to bypass the multidrug-resistance mechanism induced by tumor cells against conventional chemotherapy drugs and finally capability to produce in commercially available scale and make AMPs/ACPs as suitable candidates for the development of novel anticancer agents. Furthermore, the ease of various chemical modifications allows AMP/ACPs to be utilized alone or combined with routinely used treatments (such as peptide drug conjugates) for tumor targeting during combination therapy (12-15). Interesting features of peptides and remarkable advances in the biotechnology industry led to an increase in approved peptidebased drugs that revolutionized the pharmaceutical market. The impact of biotechnology products on pharmaceutical industries and new therapeutics can be illustrated by the fact that a remarkable percentage of recent drug approvals bv the FDA are in the biological category

(recombinant proteins and peptides, monoclonal antibodies, etc.). For example, 3 peptide-based drugs, 2 antibody-drug conjugates, 10 monoclonal antibodies, and 2 oligonucleotides were approved in 2020. The high potential of peptide or protein-based pharmaceuticals provides a clear perspective for the pharmaceutical industry, biotechnology companies, and researchers to treat disease conditions (16).

In this review article, we presented an overview of the studies focusing on AMPs/ACPs classification, mechanisms of action, and selectivity factors, and then pointed out some of the ACPs produced for targeting different types of malignancies. Also, advanced strategies described for designing therapeutic peptides, AMPs/ACPs applied in targeted therapy approaches, as those peptides engaged in clinical trials were discussed. Briefly, we focused herein on the prospect of the anticancer activity of the mentioned peptides.

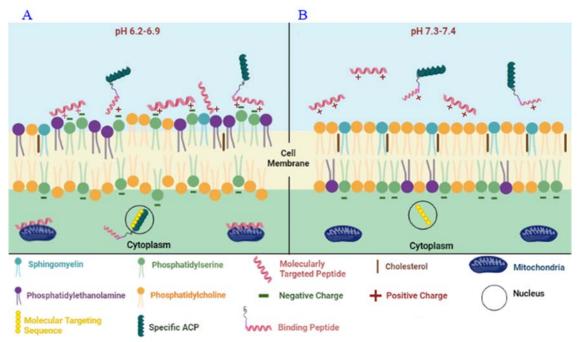


Fig. 1. Comparisons of healthy and cancerous membrane characteristics and selectivity of ACPs to bind to them. (A) Cancerous membranes and the actions of ACPs on them. Due to the negative net charge on the outer leaflet of membranes of cancer cells and the positive net charge of ACPs, ACPs can attach to the membranes and then penetrate to the cancer cell and target directly the membrane of a specific organ, causing apoptotic cell death and (B) healthy cells. The healthy cell membrane outer leaflet has a neutral net charge, which prevents ACPs from interacting. ACPs, anticancer peptides.

Classification of ACPs

Following the discovery of cecropins, several bioactive peptides with diverse bioactivities. such as immune system modulation and anti-tumor properties, were found (17). As previously stated, various types of ACPs are obtained from different organisms and categorized in numerous ways. Different models can be used for the classification of ACPs, i.e., based on their secondary structure (18), amphipathicity (19), or sources. ACPs are structurally classified into 4 different groups, including alpha (α), beta (β), alpha-beta ($\alpha\beta$), and non-alpha-beta (non- $\alpha\beta$). Amphipathicity classification is based on the hydrophobic and hydrophilic (cationic and non-cationic) features of a peptide. ACPs can be obtained from different sources, such as plants, microbes, animals, etc. (20) (Table 1).

Based on sources

Animal source

Although bioactive ACPs derived from mammal species have not yet been thoroughly studied, peptides with anti-cancer properties were found primarily in the central nervous system, digestive system, muscles, heart, immune system, bones, and skin of animals. For example, long-acting natriuretic peptides, vessel dilators, kaliuretic peptides, and atrial natriuretic peptides are natriuretic peptides secreted by the heart and have been shown to have significant anti-cancer properties in prostate (21), pancreatic adenocarcinoma (22), and breast cancer (23). A report described 4 beef-derived peptides with anti-proliferative activity that can be used for cancer treatment (24).

Peptides such as alpha-fetoprotein-derived growth inhibitory peptide, angiotensin 1-7 derived from the renin-angiotensin system, peptides obtained from anchovy protein, and an 11-amino acid peptide entitled KV11 in human apolipoprotein are samples obtained from animal proteins with potent anticancer activities (25). ACPB-3 isolated from goat liver was found to have *in vitro* anticancer activity against gastric cancer stem cells and also on a human gastric cancer cell line (BGC-823) (26). The peptide was previously found to suppress BGC-823 and CD44+ cell proliferation in a concentration-dependent manner, as well as suppress globular cell proliferation (27). ACBP-3, alone or in conjunction with cisplatin, inhibits xenograft tumor development *in vivo*, and enhances chemotherapy tolerance in a mouse model by decreasing toxicity during tests (28).

Plant source

Over 300 sequences of plant-derived peptides such as vincristine, paclitaxel, vinblastine, lentinan, camptothecin derivatives, and epipodophyllotoxin have been identified and significantly used in the development of cancer chemotherapy (25). Some plant AMPs have cytotoxic activity against mammalian cells and anticancer activity against cancer cells. Anti-cancer peptides derived from plant sources originate from both medicinal and non-medicinal herbs (29). In this respect, a study revealed that RA-V (deoxybouvardin), a natural cyclopeptide derived from the Rubia yunnanensis medicinal plant, has strong anti-tumor activity in breast cancer cells. In a preclinical study, it was found that Ganoderma lucidum poly-saccharide peptide possesses anti-tumor effects (30). Rapeseed peptide is another plant-derived bioactive peptide with anticancer properties induced by apoptosis (31). A 43amino acid peptide from soy, barley, and wheat, called lunasin, has been shown to prevent the effect of chemical carcinogens in human cells (32).

Based on the secondary structure

ACPs with a β *-pleated sheet structure*

Most ACPs with β-pleated sheets found in animals and plants have 3 disulfide bonds to connect their antiparallel β-sheets (33). Bovine lactoferrin (LfcinB) an important component of the bovine immune system is a typical sheet ACPs β-pleated (34). The half-maximal inhibitory concentration (IC50) value of LfcinB in the gastric cancer cell line (MGC803) was 32 µM. MPLfcinB6

created by linking 7 arginines to LfcinB via glycine-glycine binding sites, effectively eliminates human T-leukaemia cells with an IC50 value of 25 μ M, which is half of what it was before modification (35,36). Another peptide derived from LfcinB, known as LfcinB-P13, was discovered in another study. This peptide could improve apoptosis in the hepatocellular carcinoma cell line HepG2. Its IC50 value was 50 µg.mL⁻¹, which is better than that of LfcinB (IC50: 70 µg.mL⁻¹) (17). A human neutrophil peptide (HNP-1) is another frequent endogenous-pleated peptide. This peptide has a considerable inhibitory effect (IC50 value of 2.2 μ M) on the prostate cancer cell line PC-3 (37).

ACPs with α -helical structure

A great number of α -helical ACPs have been discovered in recent years. Alpha-helical ACPs are the most widely studied kind of ACPs, but not all have potent anti-cancer effects. Alpha-helical ACPs are less complicated than pleated sheets and have a shorter length. This peptide type is abundant in the epidermis of amphibians (38). Magainin II, for example, was the first α-helical ACP discovered in African clawed frogs (39). Magainin II has anti-cancer properties. In lung cancer cells, its IC50 was 110 g.mL⁻¹ (A549) (39, 40).

The glandular secretions of golden and green bell frogs and southern bell frogs were used to extract aurein as an α -helical peptide. Aurein has shown high inhibitory activity on T98G glioblastoma cells in various studies, with an IC50 value of 2 μ M (41). As previously stated, some ACPs have only minor anti-cancer properties. L-K6 inhibits breast cancer cells (MCF-7) with IC50 values as high as $30.2 \mu M$ (42) and has an inhibitory effect on LL37 and FK-16 colorectal cancer cells (HCT116) with IC50 values of 40 µM and 30 µM, respectively (43). Notably, although these peptides have inhibitory effects on tumor cells, it has been suggested that some ACPs have side effects, such as cytotoxicity (44).

ACPs with cyclic structure

Cyclic ACPs are circled peptides with a head-to-tail cyclization foundation or cystine

knots formed by disulfide bonds (35). Cyclic ACPs are more stable than linear ACPs, and the majority of them in clinical trials are cyclic ACPs because these peptides have a considerable inhibitory effect on cancer cells (45). Three new cyclic peptides observed in the roots and leaves of the white snake plant are Diffusa cytide 1-3.

The peptides at a concentration of 0.05 μ M significantly inhibit prostate cancer cells and can prevent cancer cells from migrating in vitro (46). H-10 is a new cyclic pentapeptide that inhibits mouse malignant melanoma cells with an IC50 value of 39.68 µM while causing no cytotoxic activity in peripheral lymphocytes (47). A study found that RA-XII, derived from Taxus yunnanensis, could prevent the growth and metastasis of colorectal tumors at an IC50 value of 5 μ M by affecting some cellular signaling pathways (48). In general. according to the results of various studies, it seems that cyclic ACPs have better anticancer and less toxicity than other ACPs. It is possible that modifying cyclic ACPs may achieve the desired results in ACP research faster.

ACPs with random coil structure

ACPs with random coils lack common secondary structures and have high concentrations of glycine and proline (49). Alloferon is an ACP with glycine-rich random coil, which is derived from insects. This ACP stimulates interferon and natural killer (NK) cell synthesis in the human and animal models (50). Alloferon has immunomodulatory and antiviral effects in people infected with human papillomavirus and herpes simplex virus, indicating that this peptide has therapeutic potential (51).

KW-WK is a peptide derivative of LFcinB18-28 created by adding the amino acids tryptophan and arginine, resulting in an irregular coil in an aqueous medium. Notably, even in high concentrations, KW-WK leads to little damage to kidney cells (20). Another peptide rich in proline arginine (PR-39) derived from neutrophils showed a strong inhibitory effect on normal embryonic kidney 293T cells. A PR-39 mutant variant, PR-35, displayed less cytotoxicity.

The data demonstrated that while cytotoxicity was reduced, biological activity was conserved. PR-39 and PR-35 showed an IC50 value of 16 g.mL⁻¹, but PR-35 had a higher cell survival rate than PR-39 (52). It seems that the impact of random coil ACPs on normal cells is much lower than that of other kinds of ACPs, but their inhibitory activity on tumor cells is also lower.

ACPs with alpha-beta $(\alpha\beta)$ and non-alpha-beta $(non-\alpha\beta)$ structures

ACPs with $\alpha\beta$ structure are a class of peptides with combined α -helix and β -sheet structures. One well-known example of the $\alpha\beta$ peptide is the human β -defensin-3, which contains 3 β -strands and a short helix in the N-terminal region (53). According to a recent report, the majority of the defensin family as well as β-defensin-3, had anticancer activity both *in vitro* and *in vivo* (54). Non- $\alpha\beta$ ACPs are a class of peptides that do not adopt welldefined α or β secondary structures. Non- $\alpha\beta$ peptides exhibiting high flexibility in aqueous solution are rich in tryptophan, proline, glycine, threonine, serine, and histidine amino acids (53). Indolicidin (ILPWKWPWWPWRR) extracted from bovine neutrophils is a peptide with a non- $\alpha\beta$ structure. The existence of tryptophan in the structure of indolicidin not only plays an important role in the anti-cancer activity but also contributes to the interaction of the peptide with the cell membrane and, finally cell-penetrating ability of the indolicidin peptide (55).

Based on amphipathicity

Glycine, lysine, and leucine are the most common amino acid residues in ACPs (56). Peptides rich in arginine and lysine are hydrophobic, positively charged, and considered cationic peptides. These peptides interact with membranes by mechanisms of snorkeling. Snorkeling is proposed to increase the hydrophobic part of the protein allowing a deeper position in the membrane and thus a stronger binding. Disrupting cell membrane integrity, interaction with cancer cells with anionic membranes, and penetrating the membrane are examples of these mechanisms (57,58). Furthermore, due to the protonation of histidine under acidic pH, histidine-containing peptides can cause cancer cytotoxicity by increasing membrane permeability (59). Although cysteine in ACPs does not play a role in cancer cell selectivity or toxicity, domains that are rich in cysteine on a variety of receptors can preserve domain structures or extracellular motifs (60). Just like glycine residues, internal prolines in ACPs are important for conformational flexibility and interaction of the peptide with the membrane (61). According to some studies, glycine and serine residues slow tumor growth and have antiproliferative effects, which are beneficial in treatment process (62). Although the methionine is a moderately hydrophobic amino acid and plays a minimal role in ACPs, it can be taken in large quantities by cancer cells. A methionine-deficient diet also produces a metabolic deficit in cancer cells by halting cell proliferation (63). In early tumors, a highly hydrophobic residue, phenylalanine, is abundant and serves as a protective amino acid (64). ACPs containing phenylalanine can also improve the affinity of peptides for attacking the cancer cell membrane (65). As stated in the previous section, tryptophan is a mildly hydrophobic amino acid that may have a role in the toxicity of ACPs, such as transactivator of transportation (TAT)-Ras GTPaseactivating protein-326 peptides and indolicidin against cancer cells (66,67). Although tyrosine does not have a role in ACP toxicity, synthetic peptides including tyrosine, phenylalanine, or proline have been shown to increase cytotoxic activity (68). The tryptophan location on ACPs is critical for their entry into malignant cells, followed by an endocytic route (69). Overall, manv investigations have shown that ACPs must have hydrophobic and cationic amino acid residues to form secondary structures that are deadly to malignant cells.

Peptide	Sequence	Structure	Cancer type	Source	Reference
AAP-H (Anthopleura anjunae anti-tumor	YVPGP	Coil	Prostate cancer	Sea anemone (<i>Anthopleura anjunae</i>)	(70,71)
Bombinin-BO1	GIGSAILSAGKSIIKGLAKGLAEHF	$Coil/\alpha$ -helix	Hepatoma cell lines	Bombina orientalis	(72,73)
Pep27	MRKEFHNVLSSGQLLADKRPARDYNRK	KEFHNVLSSGQLLADKRPARDYNRK α-helix AML-2, HL-60, Jurkat, MCF-7 and SNU- 601 cell lines		Streptococcus	(74,75)
Lactoferricin B	FKC1RRWQWRMKKLGAPSITC1VRRAF	β-sheet	Lung, tongue, esophagus, liver, and colorectal cancers	Bos taurus	(76,77)
Polybia-MP1	IDWKKLLDAAKQIL	α-helical	Bladder, prostate, and multi-resistant leukemic cancer cells	Polybia paulista	(78,79)
Pardaxin	GFFALIPKIISSPLFKTLLSAVGSALSSSGGQE	α-helical	Oral squamous cell carcinoma	Fish	(80,81)
P28	LSTAADMQGVVTDGMASGLDKDYLKPDD	Coil/a-helix	Breast cancer cell lines	Azurin from <i>Pseudomonas</i> aeruginosa	(82,83)
Bovine lactoferricin	FKC1RRWQWRMKKLGAPSITC1VRRAF	Coil/a-helix	Acute lymphoblastic T, leukemia	Bovine	(84,85)
Magainin 2	GIGKWLHSAKKFGKAFVGEIMNS	α-helix	Bladder cancer cell lines, MDA-MB-231 and M14K tumor cell lines	African clawed frog	(86,87)
Femporin-1CEa	FVDLKKIANIINSIF-NH(2)	α-helical	Breast cancer	R. chensinensis	(88)
Cecropin B	KWKVFKKIEKMGRNIRNGIVKAGPAIAVLGE AKAL	α-helix	Stomach carcinoma, acute lymphoblastic, T-leukemia cells, lung carcinoma	Hyalophora cecropia	(88,89)
R-lycosin-I	RNGIVKAGPAIAVLGE	α-helical	Lung cancer	Spider venom	(90)
NRC-03	GRRKRKWLRRIGKGVKIIGGAALDHL-NH2	Coil/a-helix	Breast cancer, multiple myeloma, leukemia	Winter flounder	(91,92)
Anoplin	GLLKRIKTLL-NH2	α-helix	Leukemia	Venom sac of the solitary wasp	(93,94)
Fachyplesin-1	KWC1FRVC2YRGIC2YRRC1R-Am	β-sheet	Melanoma cell lines	Tachypleus gigas	(95,96)
BMAP-28	GGLRSLGRKILRAWKKYGPIIVPIIRI	α-helix	U-937 lymphoma cell line, K562 leukemia cell line	Bos taurus	(97)
LL-37	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLVPRT ES	α-helix	Ovarian and breast cancer	Homo sapiens	(98,99)
Magainin 2	GIGKFLHSAKKFGKAFVGEIMNS	α-helix	Leukemia, spontaneous ovarian tumor, breast cancer	Xenopus laevis	(87,100)
Melittin	GIGAVLKVLTTGLPALISWIKRKRQQ	α-helix	Breast cancer, lung cancer	Insects (honey bee)	(101-103)
Bombinin H-BO1	IIGPVLGLVGKALGGLL	Coil/a-helix	Hepatoma cell lines	Bombina orientalis	(72,104)
HNP-1 (β-defensin)	AC1YC2RIPAC3IAGERRYGTC2IYQGRLWAFC3C 1	β-sheet	Lung carcinoma	Homo sapiens	(37,106)
BR2	RAGLQFPVGRLLRRLLR	α-helix	Cervical carcinoma, breast cancer	Buforin	(106-108
Tachyplesin I Moronecidin like peptide	KWC1FRVC2YRGIC2YRRC1R FFRNLWKGAKAAFRAGHAAWRA	β-sheet α-helix	Prostate cancer Breast cancer	<i>Tachypleus tridentatus</i> Hippocampus comes	(109) (110,111)

Table 1. Some effective ACPs and their characteristics such as sequence, structure, anticancer activity, and source.

Mode of action of ACPs

Although it is not yet known how AMPs kill tumor cells, one option is to categorize them depending on their mode of action (112). ACPs and AMPs have the same structures and physicochemical properties, but ACPs do not appear to have specific secondary structures if free in solution, they form β -plate or α -sheet structures following weak electrostatic interactions with negatively charged sites on tumor cell membranes (113). Many ACPs isolated from natural sources have been well studied and identified. For example, the HNP-1 (ACYCRIPACIAGERRYGTCIYQGALWAFC C) is an AMP with extensive activity against gram-negative and gram-positive bacteria that have been shown to have low anti-tumor activity against healthy cells (114). Aurein 1.2 is an attractive ACP, an AMP derived from the frog Litoria aurea, that has shown strong antitumor capabilities in vitro and has been found to fight against 55 distinct types of tumor cell lines while showing tiny cytotoxicity (115).

Attacking the structure of the cell membrane

The first hypothesis about the effect of ACPs was that they could break cell membranes and cause apoptosis *via* cell membrane

depolarization (116). In general, multiple models of membrane permeation, such as the "barrel-stave", "carpet", and "toroidal pore" models, have been proposed to describe the mechanism action of the mentioned peptides (117). According to one study, ACPs could cause cell death after destroying cancer cells, resulting in cytoplasm discharges.

The carpet model is the name given to this proposal (118). The majority of ACPs operate directly through this process. It is worth emphasizing that ACPs are appealing since they can target only cancer cells, as opposed to chemotherapy, which destroys healthy cells as well (119). A hybrid peptide called HPRP-A1-TAT, for example, has strong anti-cancer behavior and can decimate the cancerous cell membrane with an IC50 value of 10 µM in liver, cervical, and gastric cancers (120). Another study showed an IC50 of Temporin-La in liver cancer cells of about 11.19 µM. This ACP is extracted from the skin of a bullfrog and selectively enters and kills tumor cells, leaving healthy cells intact (121). So far, most studies on ACPs have shown good anti-tumor activity using this mechanism. However, more specialized research is needed to design them accurately (Fig. 2).

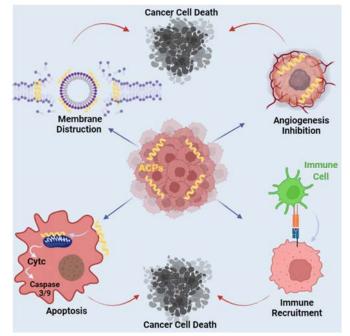


Fig. 2. Anti-tumor mechanism of ACPs. The mode of action of ACPs may include disruption of plasma/mitochondrial membranes, necrosis, apoptosis, mechanisms of mediated immunity, and angiogenesis inhibition. ACPs, anticancer peptides.

Tumor angiogenesis inhibition

Angiogenesis promotes the growth. invasion, and metastasis of solid tumors by supplying them with the nutrients and oxygen they require and removing metabolic abnormalities (122). The growth factors, including fibroblast growth factor, vascular endothelial growth factor (VEGF), epidermal growth factor, and tumor necrosis factor-alpha are involved in tumor angiogenesis. Placental growth factor, angiogenin, and platelet-derived growth factor are all examples of growth factors found in the body (123). Many peptides have shown to decrease tumor antibeen angiogenesis by interfering or interacting with growth factors and their receptors (25).

The high expression of VEGF in tumor cells could form new blood vessels. Tumor cells that have not undergone neovascularization grow slowly (124). KV11 peptide is an example of ACPs with anti-angiogenesis activity. This 11-amino-acid peptide inhibits angiogenesis by preventing microtubule formation and human umbilical vein epithelial cells (HUVEC) migration. Although the KV11 peptide did not have a considerable effect on breast cancer growth and proliferation in mice transplanted tumor models with an intense combined immune deficiency, it prevented the growth of the tumor by suppressing angiogenesis. This ACP has an IC50 of 15 µM and has no effect on HUVEC (125). Another example of ACPs with anti-angiogenesis activity is FN070315, which was isolated from the soil fungus *Penicillium* sp. It has been proven that this cyclic peptide inhibits VEGF-induced proliferation, invasion, HUVEC migration, tube formation, and neovascularization (126). The results of previous reports revealed that 2 cyclic peptides, PF1171A and PF1171C, inhibit angiogenesis by lowering the expression of the phosphorylation of VEGF receptor 2 and hypoxia-inducible factor-1 α (126) (Fig. 2). ACPs work by inhibiting neovascularization rather than killing tumor cells, so they have few side effects on normal cells. As a result, ACPs of this type have a promising clinical future.

Regulation of the immune system

LfcinB, a cationic peptide produced from lactoferrin, can boost cytokine production, hence strengthening the fight of the host against malignancies. In fact, through immune regulation, the growth of cancer can be inhibited (127). Tumor immunohistochemistry examination demonstrated that after utilizing LfcinB in cancer animals, lymphocytes increased significantly compared to untreated animals, as did tumor-infiltrating lymphocytes. It's worth noting that tumor inhibition stopped when LfcinB-induced CD3⁺ cells became tired in this study (128). A neuropeptide is MENK, which plays a role in response to tumor immune. MENK can intensify CD4+ T cell functions and secretion of cvtokines by inducing dendritic cell maturation and regulating CD8⁺ T cells. Besides, forkhead box P3 transcription factor (FOXP3) expression is inhibited, followed by reducing levels of the regulatory T cell (Treg) in vivo. All these steps end up in tumor inhibition (129). MENK also plays a role in the immune and neuroendocrine systems. It works as an immune booster and anti-tumor agent by binding to opioid receptors (130). MENK can also stop human cancer cells from proliferating by blocking cyclindependent kinase pathways (131) (Fig. 2). More research is needed to determine the possible immunomodulatory function of ACPs and how they strengthen the body's immune system against tumors.

Apoptosis

Cancer cell apoptosis is another mechanism of action by induction of α -helical ACPs through disruption of the mitochondrial membrane (132). Within eukaryotic cells, α -helical ACPs can induce mitochondrial infiltration and swelling, releasing cytochrome c (Cytc), ultimately leading to cancer cell apoptosis. The release of Cytc from the damaged mitochondria causes oligomerization of Apaf-1, activation of caspase-9, and subsequent conversion of procaspase 3 to caspase-3, which is responsible for many of the apparent symptoms of apoptosis (133,134).

A peptide derived from Meretrix could induce apoptosis, increase reactive oxygen species in the K562 cell cycle, eliminate electrical potential on the membrane surface, and degrade microcracks (135). One study confirmed that paradox induces apoptosis in the HT-1080 cell line by inhibiting caspase and disrupting the mitochondrial membrane. releasing Cytc. In the mitochondrial pathway, the induction of apoptosis in cancer cells is also associated with the death receptor pathway (136). Furthermore, it has been shown that synthetic tachyplesin conjugated to the integrin homing domain induces apoptosis of cancer cells through the aforementioned pathways. ACPs have been shown in numerous studies to cause Cytc release and stimulate apoptosis in tumor cells by wrecking the mitochondrial membrane (36). Ra-V peptide, for example, provokes apoptosis of mitochondria, which causes human breast cancer cells to die by mediating caspase signaling pathway activation, the release of Cytc, and mitochondrial membrane potential loss (36). Dolastatin 10 ACP derived from the marine mollusk Dolabella auricularia has significant cytotoxicity against various human cancer cell lines. This peptide can induce apoptosis in tumor cell lines with downregulated antiapoptotic molecule Bcl-2 (137) (Fig. 2).

Selectivity of ACPs and targeted therapy by ACPs

Regarding cell targets and selectivity of ACPs, ACPs can be classified into 2 main groups. The first group is peptides that are only active against cancer cells and microbes and do not harm healthy mammalian cells, while the second group of ACPs is peptides that do not have the ability to diagnose all 3 groups of cells (healthy, cancerous, and microbial cells) (29). Although various results have been published about experiments on the selective criteria of ACPs that kill cancer cells, their selectivity is still controversial. ACPs have been shown to generally exert their oncolytic effects by nonmembrane or membrane mechanisms (138). The mechanism of each membranolytic peptide's activity is influenced by the characteristics of ACP and the target membrane, which impact the selectivity of

many distinct differences, which contribute to the selectivity of some ACPs. The first difference is the net negative charge on the membrane, which characterizes malignant cells (139). There are several anionic molecules in the membrane of cancer cells, such as Oglycosylated mucins. heparin sulfate phosphatidylserine, and sialylated gangliosides, which give them a net negative charge while normal mammalian cell membrane is zwitterionic (29) Increasing the content of sialic acid on the membrane leads to enhancing the surface concentration of acidic groups and thus changing the membrane charge (140). The glycosylation characteristics of cancerous tissues are linked to their phenotype. Another feature of most cancer cells is that their membranes are more fluid than normal cells, which ACPs can disrupt, leading to increased permeability and potential cell death (141). ACPs can also have more contact with the microvilli in malignant cells because the cell surface area of cancer cells is much larger than that of healthy cells, so this can also be a selectivity option for ACPs. The membrane of cancer cells has a negative charge, which is also the same in bacterial cells. Thus, it can be said that the enhanced anionicity of the cytoplasmic membrane of cancerous cells and the swelling of mitochondria with Cytc release may explain the selectivity and membranolytic activity of ACPs. Various approaches to tumor focus management on addressing the angiogenesis process. Peptides inhibit the action of receptors expressed on angiogenic endothelial cells and, as a result, disrupt the establishment of the vasculature associated with a tumor (142,143). Molecularly targeted ACPs can bind, penetrate, and inhibit or destroy cancer cells at any phase of carcinogenesis or growth. As previously stated, peptides are categorized into 2 types, including a. peptides that are only effective against cancer cells and do not affect healthy cells and b. peptides that are effective against both cancer and healthy cells (144). ACPs derived from lactoferricin B, chrysophsin-1, cecropins, and magainin-2 are peptides that have only selectivity for cancer cells and not healthy cells (145). The cancer PPD database (http://crdd.osdd.net/raghava/

ACPs. Cancer and normal cells appear to have

cancerppd/) is used to predict the structure of peptides and recommend the appropriate ACP for further study (146). Furthermore, techniques that consider binary profiles, amino acid contents, and sequence-based methods are employed to target the desired cancer cells (147).

Membranolytic ACPs are synthesized from scratch utilizing designs based on helical cationic amphipathic peptide sequences (148). Anionic molecules in cancerous cells confer a net negative charge, whereas healthy cell membranes contain a neutral net charge (139). Healthy cells have high cholesterol levels in their membrane, which can prevent cationic peptides from entering through the cell fluid. Furthermore, healthy cells contain less fluid than cancer cells (149). Mastoparan-I is an α helical structure peptide and plays a role in cell swelling, cell bursting, and necrosis by interacting on the negative charge of cell surfaces of liver and prostate cancer (150). Also, an SVS-1 that is a β -sheet structure peptide breaks cell membranes in lung and breast cancer by forming pores (151). The amino acid content of ACPs is crucial in the therapy of several forms of cancer. Cationic peptides, for example, can increase the specificity of ACPs, but increasing hydrophobic peptides can decrease the degree of specificity (152). Furthermore, polycationic peptides demonstrated selectivity for acute T-cell leukemia because they have a larger membrane potential than normal tissues (153). ACPs design strategies, such as hybridization, cyclization, modification, and fragmentation, can potentially improve the therapeutic efficacy by extending the half-life time of medications in plasma, boosting activity, and minimizing drug toxicity (154).

Targeted therapy by ACPs means ACPs can bind to receptors on the cancer cell surface, allowing cell internalization (155). Therapeutic peptides are further divided into 3 types based on their biological targets, which include i. cell cycle regulation, ii. signal transduction pathways; and iii. cell death pathways (156). KLA is a tumor-penetrating ACP that promotes apoptosis. In reality, by disrupting the mitochondrial membrane, KLA causes programmed cell death (157). One of the targeted therapy approaches by ACPs is their application in the structure of fusion proteins. Fusion proteins are chimeric proteins composed of targeting and toxic moieties (107).

Denileukin diftitox (Ontak), approved by the FDA in 1999 against recurrent cutaneous T-cell lymphoma, is one of the first recombinant engineered chimeric proteins that combined interleukin-2 and diphtheria toxin. In the structure of Ontak, IL-2 and diphtheria toxin are responsible for targeting and toxic activity, respectively (158). Due to the versatile features of ACPs, they could be applied not only as tumor-targeting moieties but also as toxic or effector moieties in the structure of fusion proteins. Previously, several research groups have demonstrated that ACPs, either as targeting or toxic moiety, play a crucial role in the targeted therapy of multiple cancers (91,106,159,160). Also, a recent study used IL-24 (a pro-apoptotic cytokine) combined with p28, a tumor-specific or cell-internalizing peptide against breast cancer. The anti-tumor effects of engineered p28-IL-24 recombinant protein were investigated in vitro and in vivo. This novel fusion protein induced apoptosis and suppressed the growth of MDA-MB-231 and MCF-7 cancer cells without affecting HUVEC normal cells (159).

Clinical trials and approved ACPs

The therapeutical utility of therapeutic peptides is straightforward because only in the USA were nearly 140 clinical trials registered to evaluate the eligibility of peptides for cancer treatment. Among these peptides, ACPs appear to comprise a notable proportion of all agents entering into clinical trials (161,162). Some of the well-known ACPs at the different phases of clinical trials have been summarized in Table 2 (see website https://clinicaltrials.gov).

Peptide name	Phase	Disease	NCT number	Condition
p-28	Phase 1	Recurrent or progressive central nervous system tumors	NCT01975116	Completed
	Phase 1	Refractory solid tumors	NCT00914914	Completed
Nerofe	Phase 1	Solid tumors	NCT01690741	Completed
G250	Phases 1	Renal cell carcinoma	NCT00520533	Completed
	Phases 1 and 2	Kidney cancer	NCT00003102	Completed
Aplidine (plitidepsin)	Phase 1	Multiple myeloma	NCT02100657	Completed
	Phase 1	Advanced solid tumors lymphomas	NCT00788099	Completed
	Phase 2	Myelofibrosis	NCT01149681	Completed
MUC1	Phase 1	Non-small cell lung cancer (NSCLC) Stage III	NCT01731587	Withdrawn
LL-37	Phases 1 and 2	Melanoma	NCT02225366	Completed
iRGD	Phase 1	Acinar cell adenocarcinoma of the pancreas Duct cell adenocarcinoma of the pancreas Liver metastases Lung metastases Recurrent breast cancer Recurrent pancreatic cancer Stage IV breast cancer Stage IV pancreatic cancer	NCT01741597	Withdrawn
ATN-161	Phases 1 and 2	Brain and central nervous system tumors	NCT00352313	Completed
	Phases 2	Renal cell carcinoma	NCT00131651	Completed
LTX-315	Phase 1	Melanoma Breast cancer Head and neck cancer Lymphoma Cancer with transdermal accessible tumor Carcinoma Basal cell carcinoma	NCT01986426 NCT01058616 NCT01223209	Completed Completed Completed
	Phase 2	Skin cancer Cancer of the skin, basal cell Cancer of the skin	NCT05188729	Recruiting
LTX-315 in combination with pembrolizumab	Phase 2	Advanced melanoma	NCT04796194	Recruiting
LTX-315 and TILs	Phase 2	Soft tissue sarcoma	NCT03725605	Active, not recruiting
ANG-1005	Phase 2	Breast cancer Brain metastases Brain tumor Glioblastoma	NCT02048059 NCT01967810	Completed
	Phase 1	Advanced solid tumors with and without brain metastases	NCT00539383	Completed
		Recurrent or progressive malignant glioma	NCT00539344	Completed

Table 2. Some anticancer peptides in clinical trials, their phase, diseases, NCT number, and conditions.

Obstacles in clinical trials

A problem is the lack of selectivity of the available drugs and their consequent undesirable side effects for the patients (163). As a result, there is a need to design more selective medicines with fewer adverse effects for non-target cells. It is preferable for these novel chemicals to have distinct modes of action in relation to a specific molecule in the target cells (164). ACPs have captured the interest of several researchers due to their potential to kill or impede the growth of a wide range of bacteria and tumor cells. There are thousands of synthetic and natural peptides, many of which have anti-cancer action (165). However, only a few are now undergoing clinical testing. This is primarily owing to the numerous hurdles connected with producing these peptides into medicines, such as manufacturing costs. As a result, scientists are attempting to create new ACPs utilizing the initial restructuring of natural peptides so that the physicochemical features can be easily modified while also lowering production costs. Because certain peptides have negative effects on other healthy cells, such as being highly toxic or altering the immunological response, there are still concerns about the use of ACPs (166,167). Another critical point is the sensitivity of peptides to proteolysis, while oral administration is the preferred method of drug delivery (168). As a result, these medicines are typically administered via intravenous or intramuscular injection because feeding leads to low resistance to proteases (169). The use of a synthesis method to substitute naturally occurring amino acids with synthetic amino acids can reduce vulnerability to proteolytic degradation (170). Furthermore, determining the time of circulation, which is critical for drug efficacy, is difficult (171). Various solutions to this challenge have been offered, including the medication vectors use such of as bacteriophages (172).The use of а bacteriophage on ACPs increases targeting and enables increased activity.

technique Another great for the improvement of target specificity is to bind the ACPs to cell-penetrating peptides (CPPs). Accordingly, one study used the TAT protein of HIV as CPP to increase ACP-selectivity in cancer cells (173). The conjugation of ACPs to polymers such as polyethylene glycol has also been found to improve dynamics/pharmacokinetics by boosting penetration into desired cancer cells and allowing for additional circulation time (174). As a result, these alterations could affect the amphipathicity characteristics of therapeutic ACPs and lower their cytotoxicity against healthy cells. It also makes ACPs resistant to proteolysis while maintaining anti-cancer effects. Thus, their design and medicinal action will be improved (175).

CONCLUSION

One of the most critical issues of cancer is heterogeneity, which is a considerable obstacle to the success of cancer therapy. Although the exact mechanism of action of ACPs is still controversial, some characteristics of malignant cells make them susceptible to peptides. ACPs bind to negatively charged structures (like cancer cells) in a non-specific fashion, which are both exclusively and homogenously displayed by cancer cells. Negatively charged targets are mainly represented by phospholipids, such as PS, which are secluded in the inner side of the plasmatic membrane in normal cells, but increasing the content of PS and accordingly increasing the cancer cell's negative charge allows for the specificity of ACPs (176-178).

For the *in-silico* design of a construct from ACPs (like fusion proteins), the apoptosisinducing anticancer peptides database (ApInAPDB) could be used. ApInAPDB (http://bioinf.modares.ac.ir/software/ApInAPD B/) is a recently established database composed of about 850 apoptosis-inducing peptides and their analogues provided from previous literature, including peptides binding target or binding affinity. function, and their reported Other effectiveness as IC50. information like charge, hydrophobicity, amino acid composition, and also prediction of secondary structure using different algorithms are accessible in the mentioned database (179). I-TASSER and RAMPAGE are other wellknown web servers that could be used to design ACPs and fusion proteins. To predict 3dimensional structures and the evaluation of molecular dynamic behaviors, MODELLER and GROMACS software are accessible, respectively (180).

In recent years, peptide design has benefited tremendously from advancements in artificial intelligence algorithms. These algorithms have greatly facilitated and accelerated the process of peptide discovery and optimization. By leveraging complex computational models, machine learning techniques, and large datasets, artificial intelligence algorithms can efficiently analyze the vast space of peptide sequences and structures, predicting their potential anti-cancer activity and identifying optimal candidates for further experimental validation. Moreover, these algorithms can also take into account various physicochemical and structural properties of peptides, enabling the design of molecules with enhanced stability, selectivity, and bioavailability. The integration of artificial intelligence algorithms in peptide design has not only improved the efficiency of the discovery process but has also opened up new avenues for the rational design of novel peptide-based therapies with improved efficacy and specificity. As such, the exploration and application of artificial intelligence algorithms for peptide design hold great promise for advancing the field of anti-cancer peptide therapeutics (181,182).

Although it was reported that the specificity of the ACPs is better than that of chemotherapy drugs, in some cases, especially in the case of synthetic ACPs, lower specificity toward cancer cells might be observed. The lower specificity of ACPs generally refers to their physicochemical properties, such as charge, hydrophobicity, and structure. Investigations have revealed that ACPs with high hydrophobicity and a positive net charge selectively kill cancer cells by interacting with anionic cell membrane components of cancer cells (183,184). So, rational design of well ACPs as as manipulation of physicochemical properties are 2 kev parameters to enhance specificity. For example, in a study by Fu et al., the TAT-KLA peptide was conjugated to the BRBP1 peptide, which was previously identified for its affinity toward the MDA-MB-231 cell line, to enhance its specificity against tumor cells. (185). In another study, the specificity of the HPRP-A1 peptide compared with the HPRP-A1-TAT peptide was evaluated. Between HPRP-A1 alone and HPRP-A1-TAT, the latter has higher positive charges and may have more chances to interact with the anionic surface of cancer cells (173).

cancer Besides heterogenicity and specificity, some other issues still exist in cancer treatment by ACPs. The challenges for using ACPs in cancer therapy are the poor bioavailability, immune response to treatments, toxicity of the peptides, and the costinefficiency of the approaches. Due to the peptide nature of ACPs, proteolytic degradation is a major threat to the potency of peptide-based drugs, which decreases their bioavailability and limits the systemic delivery potential of ACPs. To overcome lower bioavailability as well as immune response issues, various delivery systems (encapsulation of ACPs in liposomes, polymer nanoparticles, or quantum dots) were used (186).

Since most existing anti-cancer medications attack all rapidly dividing cells, existing cancer therapies have numerous adverse side effects. Although more research on the specific mechanism of ACPs on cancer cells is needed, several studies have demonstrated that many ACPs are capable of targeting cancer cells while avoiding damage to healthy cells. As a result, ACP therapy has an impact on molecular targets by binding anticancer medications to the target cell and stimulating biological processes (143). The emergence of highly cationic anticancer peptides as potent anticancer agents has opened new doors in cancer therapeutics. These peptides exhibit selectivity towards cancer cells while sparing normal mammalian cells, making them ideal candidates for targeted and less-toxic cancer treatments. As research in this field progresses, it is expected that more AMPs will be identified and developed for targeting the different types of malignancies. One promising aspect is the combination of ACPs with conventional medications, which may lead to synergistic effects and improved treatment outcomes. The ability of ACPs to penetrate tumors effectively due to their strong tumor penetration and solubility further adds to their potential as successful cancer treatments. In addition to preclinical studies, the translation of promising ACPs into clinical trials is a crucial step toward their development as cancer therapeutics. Current clinical trials evaluating the safety and efficacy of these peptides will provide valuable insights into their potential use in clinical practice (187-189).

ACPs possess unique pharmacokinetic properties that make them an ideal option for cancer therapy. These peptides are small in size, allowing for easy penetration into tumor tissues and cellular membranes. They exhibit high selectivity towards cancer cells, minimizing off-target effects and reducing toxicity to healthy tissues. Additionally, anti-cancer peptides have a short plasma half-life, which ensures rapid clearance from the body and reduces the risk of accumulation and adverse Furthermore, reactions. their inherent biodegradability and low immunogenicity make them suitable for repeated administration. These pharmacokinetic properties of anticancer peptides enhance their therapeutic efficacy and hold promise for the development of targeted and personalized cancer treatments (190,191). One intriguing aspect of targeted therapy utilizing ACPs is their potential application in cancer imaging. ACPs have gained attention for their potential in cancer imaging, a targeted therapy approach. ACPs can be labeled with imaging agents, such as fluorescent dyes, radioactive isotopes, or nanoparticles, which enable their visualization in real time through imaging techniques like fluorescence imaging, positron emission tomography, and magnetic resonance imaging. This opens up possibilities for early detection, precise diagnosis, and monitoring of tumors. ACPs selectively bind to cancer cells, making them useful molecular probes for tumorspecific imaging. This targeted imaging approach can assist in detecting small or hidden tumors, monitoring treatment response, and guiding surgical resection. Additionally, cancer imaging with ACPs is non-invasive. minimizing patient discomfort while providing valuable information for personalized cancer management (192,193).

The future of peptides in cancer treatment looks promising, thanks to the advancements in personalized medicine, peptide engineering, drug delivery systems, and combination therapies. Personalized approaches using omics data can identify specific molecular targets, allowing peptides to deliver therapeutic payloads directly to cancer cells. Peptide engineering techniques enhance stability and efficacy, while novel drug delivery systems enable efficient tumor targeting. Combining peptides with other therapies can lead to synergistic effects and improved outcomes. Artificial intelligence algorithms expedite peptide design, leading to the discovery of novel sequences with enhanced properties. Together, these advancements hold the potential for highly targeted, effective, and personalized peptide-based cancer therapies.

Overall, the rapid progress in the understanding and development of ACPs as anticancer agents brings hope for improved cancer treatments with enhanced selectivity, reduced toxicity, and increased efficacy. Continued research and clinical trials will pave the way for the integration of ACPs into standard cancer treatment regimens, ultimately improving patient outcomes and reducing the global burden of cancer (194).

To that purpose, natural and synthesized peptides are novel cancer-fighting agents. Many ACPs have anti-apoptotic and antiproliferative properties in various types of cancer cells, both *in vitro* and *in vivo*, which is why they have been tested in clinical trials for cancer treatment. In addition, clinical research believes that ACPs will boost cancer medications to prevent new instances of cancer and its related death cases.

Acknowledgments

This work was financially supported by the Research Deputy of Isfahan University of Medical Sciences *via* Grant No. 1400494. We also thank the CinnaGen Medical Biotechnology Research Center.

Conflict of interest statement

All authors declared no conflict of interest in this study.

Authors' contributions

A. Jahanian-Najafabadi contributed the initial idea and provided supervision throughout the research process; E Khodamoradi was responsible for preparing the references and collaborated on the preparation of figures and tables; R. Ghavimi and S. Mahmoudi took the lead in drafting the original manuscript; A. Jahanian-Najafabadi and M. Mohammadi reviewed and edited the manuscript: M. Mohammadi and E. Khodamoradi assisted in the preparation of figures and tables. All authors read and approved the finalized version of the manuscript.

REFERENCES

- Blackadar CB. Historical review of the causes of cancer. World J Clin Oncol. 2016;7(1):54-86. DOI: 10.5306/wjco.v7.i1.54.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, *et al.* Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209-249. DOI: 10.3322/caac.21660.
- Momenzadeh N, Hajian S, Shabankare A, Ghavimi R, Kabiri-Samani S, Kabiri H, *et al.* Photothermic therapy with cuttlefish ink-based nanoparticles in combination with anti-OX40 mAb achieve remission of triple-negative breast cancer. Int Immunopharmacol. 2023;115:109622,1-2. DOI: 10.1016/j.intimp.2022.109622.
- Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. CA Cancer J Clin. 2024;74(1):12-49. DOI: 10.3322/caac.21820.

- Thun MJ, DeLancey JO, Center MM, Jemal A, Ward EM. The global burden of cancer: priorities for prevention. Carcinogenesis. 2010;31(1):100-110. DOI: 10.1093/carcin/bgp263.
- Abbas Z, Rehman S. An overview of cancer treatment modalities. Neoplasm. 2018;1:139-157. DOI: 10.5772/intechopen.76558.
- Falzone L, Salomone S, Libra M. Evolution of cancer pharmacological treatments at the turn of the third millennium. Front Pharmacol. 2018;9:1300,1-26. DOI: 10.3389/fphar.2018.01300.
- Pucci C, Martinelli C, Ciofani G. Innovative approaches for cancer treatment: current perspectives and new challenges. Ecancermedicalscience. 2019;13:961,1-26. DOI: 10.3332/ecancer.2019.961.
- Bukowski K, Kciuk M, Kontek R. Mechanisms of multidrug resistance in cancer chemotherapy. Int J Mol Sci. 2020;21(9):3233,1-24. DOI: 10.3390/ijms21093233.
- 10. Padma VV. An overview of targeted cancer therapy. BioMedicine. 2015;5(4):19,1-6. DOI: 10.7603/s40681-015-0019-4.
- Chari RV. Targeted cancer therapy: conferring specificity to cytotoxic drugs. Acc Chem Res. 2008;41(1):98-107. DOI: 10.1021/ar700108g.
- Liscano Y, Oñate-Garzón J, Delgado JP. Peptides with dual antimicrobial-anticancer activity: strategies to overcome peptide limitations and rational design of anticancer peptides. Molecules. 2020;25(18): 4245,1-20.

DOI: 10.3390/molecules25184245.

- Taheri B, Mohammadi M, Nabipour I, Momenzadeh N, Roozbehani M. Identification of novel antimicrobial peptide from Asian sea bass (lates calcarifer) by *in silico* and activity characterization. PLoS One. 2018;13(10):e0206578,1-22. DOI: 10.1371/journal.pone.0206578.
- 14. Ghavimi R, Mohammadi E, Akbari V, Shafiee F, Jahanian-Najafabadi A. *In silico* design of two novel fusion proteins, p28-IL-24 and p28-M4, targeted to breast cancer cells. Res Pharm Sci. 2020;15(2): 200-208.

DOI: 10.4103/1735-5362.283820.

- 15. Tolos Vasii AM, Moisa C, Dochia M, Popa C, Copolovici L, Copolovici DM. Anticancer potential of antimicrobial peptides: focus on buforins. Polymers. 2024;16(6):728,1-14. DOI: 10.3390/polym16060728.
- 16. de la Torre BG, Albericio F. The pharmaceutical industry in 2020. An analysis of FDA drug approvals from the perspective of molecules. Molecules. 2021;26(3):627,1-14. DOI: 10.3390/molecules26030627.
- Qu B, Yuan J, Liu X, Zhang S, Ma X, Lu L. Anticancer activities of natural antimicrobial peptides from animals. Front Microbiol. 2024;14:1321386,1-17. DOI: 10.3389/fmicb.2023.1321386.
- 18. Li Y, Xiang Q, Zhang Q, Huang Y, Su Z. Overview on the recent study of antimicrobial peptides: origins, functions, relative mechanisms and application. Peptides. 2012;37(2):207-225. DOI: 10.1016/j.peptides.2012.07.001.

19. Huang YB, Wang XF, Wang HY, Liu Y, Chen Y. Studies on mechanism of action of anticancer peptides by modulation of hydrophobicity within a defined structural framework. Mol Cancer Ther. 2011;10(3):416-46.

DOI: 10.1158/1535-7163.MCT-10-0811.

- 20. Wang H, Li Y, Hou J, Jiang C, Li D, Lu J, *et al.* Biological activity and stability of new peptide derivative KW-WK from bovine lactoferricin. Food Sci. 2018;39(20):57-62. DOI: 10.7506/spkx1002-6630-201820009.
- 21. Vesely B, Alli A, Song S, Gower Jr W, Sanchez-Ramos J, Vesely D. Four peptide hormones' specific decrease (up to 97%) of human prostate carcinoma cells. Eur J Clin Invest. 2005;35(11): 700-710.

DOI: 10.1111/j.1365-2362.2005.01569.x.

- 22. Skelton IV WP, Skelton M, Vesely DL. Cardiac hormones are potent inhibitors of secreted frizzledrelated protein-3 in human cancer cells. Exp Ther Med. 2013;5(2):475-478. DOI: 10.3892/etm.2012.806.
- 23. Vesely B, Song S, Sanchez-Ramos J, Fitz S, Solivan S, R. Gower Jr W, *et al.* Four peptide hormones decrease the number of human breast adenocarcinoma cells. Eur J Clin Invest. 2005;35(1):60-69.

DOI: 10.1111/j.1365-2362.2005.01444.x.

- 24. Jang A, Jo C, Kang KS, Lee M. Antimicrobial and human cancer cell cytotoxic effect of synthetic angiotensin-converting enzyme (ACE) inhibitory peptides. Food Chem. 2008;107(1):327-336. DOI: 10.1016/j.foodchem.2007.08.036.
- 25. Wu D, Gao Y, Qi Y, Chen L, Ma Y, Li Y. Peptidebased cancer therapy: opportunity and challenge. Cancer Lett. 2014;351(1):13-22. DOI: 10.1016/j.canlet.2014.05.002.
- 26. Su L, Xu G, Shen J, Tuo Y, Zhang X, Jia S, *et al.* Anticancer bioactive peptide suppresses human gastric cancer growth through modulation of apoptosis and the cell cycle. Oncol Rep. 2010;23(1):3-9. PMID: 19956858.
- 27. Yu L, Yang L, An W, Su X. Anticancer bioactive peptide-3 inhibits human gastric cancer growth by suppressing gastric cancer stem cells. J Cell Biochem. 2014;115(4):697-711. DOI: 10.1002/jcb.24711.
- 28. Su X, Dong C, Zhang J, Su L, Wang X, Cui H, et al. Combination therapy of anti-cancer bioactive peptide with cisplatin decreases chemotherapy dosing and toxicity to improve the quality of life in xenograft nude mice bearing human gastric cancer. Cell Biosci. 2014;4(1):7,1-13.

DOI: 10.1186/2045-3701-4-7.

- Hoskin DW, Ramamoorthy A. Studies on anticancer activities of antimicrobial peptides. Biochim-Biophys Acta. 2008;1778(2):357-375. DOI: 10.1016/j.bbamem.2007.11.008.
- 30. Papo N, Shai Y. Host defense peptides as new weapons in cancer treatment. Cell Mol Life Sci. 2005;62(7-8):784-790. DOI: 10.1007/s00018-005-4560-2.

31. Fan H, Liu H, Zhang Y, Zhang S, Liu T, Wang D. Review on plant-derived bioactive peptides: biological activities, mechanism of action and utilizations in food development. J Future Foods. 2022;2(2):143–159.

DOI: 10.1016/j.jfutfo.2022.03.003.

- 32. Hernandez-Ledesma B, Hsieh CC, Ben O. Lunasin, a novel seed peptide for cancer prevention. Peptides. 2009;30(2):426-430. DOI: 10.1016/j.peptides.2008.11.002.
- 33. Brock K, Talley K, Coley K, Kundrotas P, Alexov E. Optimization of electrostatic interactions in proteinprotein complexes. Biophys J. 2007;93(10):3340-3352.

DOI: 10.1529/biophysj.107.112367.

- 34. Bellamy W, Takase M, Yamauchi K, Wakabayashi H, Kawase K, Tomita M. Identification of the bactericidal domain of lactoferrin. Biochim-Biophys Acta. 1992;1121(1-2):130-136. DOI: 10.1016/0167-4838(92)90346-F.
- 35. Xie M, Liu D, Yang Y. Anti-cancer peptides: classification, mechanism of action, reconstruction and modification. Open Biol. 2020;10(7):200004,1-10.

DOI: 10.1098/rsob.200004.

36. Hilchie AL, Vale R, Zemlak TS, Hoskin DW. Generation of a hematologic malignancy-selective membranolytic peptide from the antimicrobial core (RRWQWR) of bovine lactoferricin. Exp Mol Pathol. 2013;95(2):192-198. DOL:10.1016/j.

DOI: 10.1016/j.yexmp.2013.07.006.

37. Gaspar D, Freire JM, Pacheco TR, Barata JT, Castanho MA. Apoptotic human neutrophil peptide-1 anti-tumor activity revealed by cellular biomechanics. Biochim Biophys Acta. 2015;1853(2):308-316.

DOI: 10.1016/j.bbamcr.2014.11.006.

- 38. Dong TT, Tian ZG, Wang JH. The structural parameters-functional activity relationship of alphahelical antimicrobial peptides. Chin Biotechnol. 2007;27(9):116-119. DOI: 10.1109/5.771073.
- 39. Khavani M, Mehranfar A, Mofrad MRK. Antimicrobial peptide interactions with bacterial cell membranes. J Biomol Struct Dyn. 2024:1-14. DOI: 10.1080/07391102.2024.2304683.
- 40. Kim MK, Kang NH, Ko SJ, Park J, Park E, Shin DW, et al. Antibacterial and antibiofilm activity and mode of action of magainin 2 against drug-resistant *Acinetobacter baumannii*. Int J Mol Sci. 2018;19(10):3041,1-14. DOI: 10.3390/ijms19103041.
- 41. Dennison SR, Harris F, Phoenix DA. The interactions of aurein 1.2 with cancer cell membranes. Biophys Chem. 2007;127(1-2):78-83. DOI: 10.1016/j.bpc.2006.12.009.
- 42. Wang C, Dong S, Zhang L, Zhao Y, Huang L, Gong X, *et al.* Cell surface binding, uptaking and anticancer activity of L-K6, a lysine/leucine-rich peptide, on human breast cancer MCF-7 cells. Sci Rep. 2017;7(1):8293,1-13. DOI: 10.1038/s41598-017-08963-2.

43. Ren SX, Shen J, Cheng AS, Lu L, Chan RL, Li ZJ, et al. FK-16 derived from the anticancer peptide LL-37 induces caspase-independent apoptosis and autophagic cell death in colon cancer cells. PLoS One. 2013;8(5):e63641,1-9. DOI: 10.1371/journal.pone.0063641

DOI: 10.1371/journal.pone.0063641.

- 44. Tran TH, Le TH, Tran TTP. The potential effect of endogenous antimicrobial peptides in cancer immunotherapy and prevention. J Pept Sci. 2025;31(2):e3664. DOI: 10.1002/psc.3664.
- 45. Lee Y, Phat C, Hong SC. Structural diversity of marine cyclic peptides and their molecular mechanisms for anticancer, antibacterial, antifungal, and other clinical applications. Peptides. 2017;95:94-105.

DOI: 10.1016/j.peptides.2017.06.002.

- 46. Hu E, Wang D, Chen J, Tao X. Novel cyclotides from *Hedyotis diffusa* induce apoptosis and inhibit proliferation and migration of prostate cancer cells. Int J Clin Exp Med. 2015;8(3):4059-4065. PMID: 26064310.
- 47. Zhang Y, Fu J, Zhang Z, Qin H. miR-486-5p regulates the migration and invasion of colorectal cancer cells through targeting PIK3R1. Oncol Lett. 2018;15(5):7243-7248. DOI: 10.3892/ol.2018.8233.
- 48. Wang Y, Guo D, He J, Song L, Chen H, Zhang Z, et al. Inhibition of fatty acid synthesis arrests colorectal neoplasm growth and metastasis: anti-cancer therapeutical effects of natural cyclopeptide RA-XII. Biochem Biophys Res Commun. 2019;512(4):819-824.

DOI: 10.1016/j.bbrc.2019.03.088.

49. Veldhuizen EJ, Schneider VA, Agustiandari H, Van Dijk A, Tjeerdsma-van Bokhoven JL, *et al.* Antimicrobial and immunomodulatory activities of PR-39 derived peptides. PLoS One. 2014;9(4):e95939,1-7.

DOI: 10.1371/journal.pone.0095939.

50. Bae S, Oh K, Kim H, Kim Y, Kim HR, Hwang YI, *et al.* The effect of alloferon on the enhancement of NK cell cytotoxicity against cancer via the up-regulation of perforin/granzyme B secretion. Immunobiology. 2013;218(8):1026-1033.

DOI: 10.1016/j.imbio.2012.12.002.

- 51. Appiah C, Chen S, Retyunskiy V, Tzeng C, Zhao Y. Study of alloferon, a novel immunomodulatory antimicrobial peptide (AMP), and its analogues. Front Pharmacol. 2024;15:1359261,1-16. DOI: 10.3389/fphar.2024.1359261.
- 52. Jeon H, Le MT, Ahn B, Cho HS, Le VCQ, Yum J, *et al.* Copy number variation of PR-39 cathelicidin, and identification of PR-35, a natural variant of PR-39 with reduced mammalian cytotoxicity. Gene. 2019;692:88-93.
- DOI: 10.1016/j.gene.2018.12.065.
- 53. Yan J, Cai J, Zhang B, Wang Y, Wong DF, Siu SW. Recent progress in the discovery and design of antimicrobial peptides using traditional machine learning and deep learning. Antibiotics. 2022;11(10):1451,1-30. EQU: 10.2220/1011101451

DOI: 10.3390/antibiotics11101451.

- 54. Adyns L, Proost P, Struyf S. Role of defensins in tumor biology. Int J Mol Sci. 2023;24(6):5268,1-16. DOI: 10.3390/ijms24065268.
- 55. Tripathi AK, Vishwanatha JK. Role of anti-cancer peptides as immunomodulatory agents: potential and design strategy. Pharmaceutics. 2022;14(12):2686,1-20.

DOI: 10.3390/pharmaceutics14122686.

- 56. Shoombuatong W, Schaduangrat N, Nantasenamat C. Unraveling the bioactivity of anticancer peptides as deduced from machine learning. EXCLI J. 2018;17:734-752. DOI: 10.17179/excli2018-1447.
- 57. Dai Y, Cai X, Shi W, Bi X, Su X, Pan M, et al. Proapoptotic cationic host defense peptides rich in lysine or arginine to reverse drug resistance by disrupting tumor cell membrane. Amino acids. 2017;49(9):1601-1610.

DOI: 10.1007/s00726-017-2453-y.

- 58. Strandberg E, Killian JA. Snorkeling of lysine side chains in transmembrane helices: how easy can it get? FEBS Lett. 2003;544(1-3):69-73. DOI: 10.1016/S0014-5793(03)00475-7.
- 59. Yamaguchi Y, Yamamoto K, Sato Y, Inoue S, Morinaga T, Hirano E. Combination of aspartic acid and glutamic acid inhibits tumor cell proliferation. Biomed Res. 2016;37(2):153-159. DOI: 10.2220/biomedres.37.153.
- 60. Oancea E, Teruel MN, Quest AF, Meyer T. Green fluorescent protein (GFP)-tagged cysteine-rich domains from protein kinase C as fluorescent indicators for diacylglycerol signaling in living cells. J Cell Biol. 1998;140(3):485-498. DOI: 10.1083/jcb.140.3.485.
- 61. Shamova O, Orlov D, Stegemann C, Czihal P, Hoffmann R, Brogden K, *et al.* A novel proline-rich antimicrobial peptide from goat leukocytes. Int J Pep Prot Res. 2009;15:107-119. DOI: 10.1007/s10989-009-9170-7.
- 62. Maddocks OD, Athineos D, Cheung EC, Lee P, Zhang T, van den Broek NJ, *et al.* Modulating the therapeutic response of tumours to dietary serine and glycine starvation. Nature. 2017;544(7650):372-376. DOI: 10.1038/nature22056.
- 63. Kawaguchi K, Han Q, Li S, Tan Y, Igarashi K, Kiyuna T, *et al.* Targeting methionine with oral recombinant methioninase (o-rMETase) arrests a patient-derived orthotopic xenograft (PDOX) model of BRAF-V600E mutant melanoma: implications for chronic clinical cancer therapy and prevention. Cell Cycle. 2018;17(3):356-361.

DOI: 10.1080/15384101.2017.1405195.

- 64. Gueron G, Anselmino N, Chiarella P, Ortiz EG, Lage Vickers S, Paez AV, *et al.* Game-changing restraint of Ros-damaged phenylalanine, upon tumor metastasis. Cell Death Dis. 2018;9(2):1-15. DOI: 10.1038/s41419-017-0147-8.
- 65. Dennison SR, Whittaker M, Harris F, Phoenix DA. Anticancer α -helical peptides and structure/function relationships underpinning their interactions with tumour cell membranes. Curr Protein Pept Sci. 2006;7(6):487-499.

DOI: 10.2174/138920306779025611.

66. Marchand C, Krajewski K, Lee HF, Antony S, Johnson AA, Amin R, *et al.* Covalent binding of the natural antimicrobial peptide indolicidin to DNA abasic sites. Nucleic Acids Res. 2006;34(18):5157-5165.

DOI: 10.1093/nar/gkl667.

- 67. Barras D, Chevalier N, Zoete V, Dempsey R, Lapouge K, Olayioye MA, *et al.* A WXW motif is required for the anticancer activity of the TAT-RasGAP317–326 peptide. J Biol Chem. 2014;289(34):23701-23711. DOI: 10.1074/jbc.M114.576272.
- 68. Ahmaditaba MA, Shahosseini S, Daraei B, Zarghi A, Houshdar Tehrani MH. Design, synthesis, and biological evaluation of new peptide analogues as selective COX-2 inhibitors. Arch Pharm. 2017;350(10):1700158,1-9. DOI: 10.1002/ardp.201700158.
- 69. Bhunia D, Mondal P, Das G, Saha A, Sengupta P, Jana J, *et al.* Spatial position regulates power of tryptophan: discovery of a major-groove-specific nuclear-localizing, cell-penetrating tetrapeptide. J Am Chem Soc. 2018;140(5):1697-1714. DOI: 10.1021/jacs.7b10254.
- 70. Wu ZZ, Ding GF, Huang FF, Yang ZS, Yu FM, Tang YP, *et al.* Anticancer activity of anthopleura anjunae oligopeptides in prostate cancer DU-145 cells. Mar Drugs. 2018;16(4):125,1-15. DOI: 10.3390/md16040125.
- 71. Zhang QT, Liu ZD, Wang Z, Wang T, Wang N, Wang N, *et al.* Recent advances in small peptides of marine origin in cancer therapy. Mar drugs. 2021;19(2):115,1-29. DOI: 10.3390/md19020115.
- 72. Tornesello AL, Borrelli A, Buonaguro L, Buonaguro FM, Tornesello ML. Antimicrobial peptides as anticancer agents: functional properties and biological activities. Molecules. 2020;25(12):2850,1-25.

DOI: 10.3390/molecules25122850.

- 73. Hou X, Du Q, Li R, Zhou M, Wang H, Wang L, et al. Feleucin-BO 1: a novel antimicrobial non-apeptide amide from the skin secretion of the toad, *Bombina* orientalis, and design of a potent broad-spectrum synthetic analogue, feleucin-K 3. Chem Biol Drug Des. 2015;85(3):259-267. DOI: 10.1111/cbdd.12396.
- 74. Lee DG, Hahm KS, Park Y, Kim HY, Lee W, Lim SC, *et al.* Functional and structural characteristics of anticancer peptide Pep27 analogues. Cancer Cell Int. 2005;5:21,1-14.

DOI: 10.1186/1475-2867-5-21.

- 75. Sung WS, Park Y, Choi CH, Hahm KS, Lee DG. Mode of antibacterial action of a signal peptide, Pep27 from Streptococcus pneumoniae. Biochem Biophys Res Commun. 2007;363(3):806-810. DOI: 10.1016/j.bbrc.2007.09.041.
- 76. Hwang PM, Zhou N, Shan X, Arrowsmith CH, Vogel HJ. Three-dimensional solution structure of lactoferricin B, an antimicrobial peptide derived from bovine lactoferrin. Biochemistry. 1998;37(12):4288-4298.

DOI: 10.1021/bi972323m.

- 77. Cutone A, Rosa L, Ianiro G, Lepanto MS, Bonaccorsi di Patti MC, Valenti P, et al. Lactoferrin's anti-cancer properties: safety, selectivity, and wide range of action. Biomolecules. 2020;10(3):456,1-26. DOI: 10.3390/biom10030456.
- 78. Alvares DS, Wilke N, Neto JR, Fanani ML. The insertion of Polybia-MP1 peptide into phospholipid monolayers is regulated by its anionic nature and phase state. Chem Phys Lipids. 2017;207(Pt A):38-48

DOI: 10.1016/j.chemphyslip.2017.08.001.

79. Alvares DS, Fanani ML, Neto JR, Wilke N. The interfacial properties of the peptide Polybia-MP1 and its interaction with DPPC are modulated by lateral electrostatic attractions. Biochem Biophys Acta. 2016;1858(2):393-402. DOI: 10.1016/j.bbamem.2015.12.010.

- 80. Han Y, Cui Z, Li YH, Hsu WH, Lee BH. In vitro and in vivo anticancer activity of pardaxin against proliferation and growth of oral squamous cell carcinoma. Mar Drugs. 2015;14(1):2,1-12. DOI: 10.3390/md14010002
- 81. Shinde SD, Atpadkar P, Swain P, Apparao CV, Sandhya V, Sahu B. Peptide and protein in therapeutics. In: Jain A and Malik S, editors. Peptide and protein drug delivery using polysaccharides. 1st edition. The Netherlands: Elsevier; 2024. p. 1-24. DOI: 10.1016/B978-0-443-18925-8.00007-6.
- 82. Soleimani M, Mahnam K, Mirmohammad-Sadeghi H, Sadeghi-Aliabadi H, Jahanian-Najafabadi A. Theoretical design of a new chimeric protein for the treatment of breast cancer. Res Pharm Sci. 2016;11(3):187-199. PMID: 27499788.
- 83. Soleimani M, Mirmohammad-Sadeghi H, Sadeghi-Aliabadi H, Jahanian-Najafabadi A. Expression and purification of toxic anti-breast cancer p28-NRC chimeric protein. Adv Biomed Sci. 2016;5:70,1-7. DOI: 10.4103/2277-9175.180639.
- 84. Furlong SJ, Ridgway ND, Hoskin DW. Modulation of ceramide metabolism in T-leukemia cell lines potentiates apoptosis induced by the cationic antimicrobial peptide bovine lactoferricin. Int J Oncol. 2008;32(3):537-544. PMID: 18292930.
- 85. Richardson A, de Antueno R, Duncan R, Hoskin DW. Intracellular delivery of bovine lactoferricin's antimicrobial core (RRWOWR) kills T-leukemia cells. Biochem **Biophys** Res Commun. 2009;388(4):736-741. DOI: 10.1016/j.bbrc.2009.08.083.
- 86. Lehmann J, Retz M, Sidhu SS, Suttmann H, Sell M, Paulsen F, et al. Antitumor activity of the antimicrobial peptide magainin II against bladder cancer cell lines. Eur Urol. 2006;50(1):141-147. DOI: 10.1016/j.eururo.2005.12.043.
- 87. Anghel R, Jitaru D, Bădescu L, Bădescu M, Ciocoiu M. The cytotoxic effect of magainin II on the MDA-MB-231 and M14K tumour cell lines. Biomed Res Int. 2013;2013:1-11. DOI: 10.1155/2013/831709.
- 88. Wang C, Li HB, Li S, Tian LL, Shang DJ. Antitumor effects and cell selectivity of temporin-1CEa, an

antimicrobial peptide from the skin secretions of the Chinese brown frog (Rana chensinensis). Biochimie. 2012;94(2):434-441.

DOI: 10.1016/j.biochi.2011.08.011.

- 89. Anghel R, Jitaru D, Badescu L, Ciocoiu M, Badescu M. The cytotoxic effect of Cecropin A and Cecropin B on the MDA-MB-231 and M14K tumour cell lines. J Biomed Sci Eng. 2014;7(8):1-13. DOI: 10.4236/jbise.2014.78052.
- 90. Rádis-Baptista G. Cell-penetrating peptides derived from animal venoms and toxins. Toxins. 2021;13(2):147,1-25. DOI: 10.3390/toxins13020147.
- 91. Soleimani M, Sadeghi HM, Jahanian-Najafabadi A.
- A Bi-functional targeted P28-NRC chimeric protein with enhanced cytotoxic effects on breast cancer cell lines. Iran J Pharm Res. 2019;18(2):735-744. DOI: 10.22037/ijpr.2019.2392.
- 92. Hou D, Hu F, Mao Y, Yan L, Zhang Y, Zheng Z, et al. Cationic antimicrobial peptide NRC-03 induces oral squamous cell carcinoma cell apoptosis via CypD-mPTP axis-mediated mitochondrial oxidative stress. Redox Biol. 2022;54:102355,1-19. DOI: 10.1016/j.redox.2022.102355
- 93. Zhu LN, Fu CY, Zhang SF, Chen W, Jin YT, Zhao FK. Novel cytotoxic exhibition mode of antimicrobial peptide anoplin in MEL cells, the cell line of murine Friend leukemia virus-induced leukemic cells. J Pept Sci. 2013;19(9):566-574.
- DOI: 10.1002/psc.2533.
- 94. Konno K, Hisada M, Fontana R, Lorenzi CC, Naoki H, Itagaki Y, et al. Anoplin, a novel antimicrobial peptide from the venom of the solitary wasp Anoplius samariensis. Biochem Biophys Acta. 2001;1550(1):70-80.

DOI: 10.1016/S0167-4838(01)00271-0.

- 95. Vernen F, Harvey PJ, Dias SA, Veiga AS, Huang Y-H, Craik DJ, et al. Characterization of tachyplesin peptides and their cyclized analogues to improve antimicrobial and anticancer properties. Int J Mol Sci. 2019;20(17):4184,1-25. DOI: 10.3390/ijms20174184.
- 96. Lee LF, Mariappan V, Vellasamy KM, Lee VS, Vadivelu J. Antimicrobial activity of tachyplesin 1 against Burkholderia pseudomallei: an in vitro and in silico approach. PeerJ. 2016;4:e2468,1-29. DOI: 10.7717/peerj.2468.
- 97. Risso A, Braidot E, Sordano MC, Vianello A, Macrì F, Skerlavaj B, et al. BMAP-28, an antibiotic peptide of innate immunity, induces cell death through opening of the mitochondrial permeability transition pore. Mol Cell Biol. 2002;22(6):1926-1935. DOI: 10.1128/MCB.22.6.1926-1935.2002.
- 98. Nagaoka I, Suzuki K, Niyonsaba F, Tamura H, Hirata M. Modulation of neutrophil apoptosis by antimicrobial peptides. ISRN Microbiol. 2012;2012,1-12. DOI: 10.5402/2012/345791.
- 99. Bhattacharjya S, Zhang Z, Ramamoorthy A. LL-37: structures, antimicrobial activity, and influence on amyloid-related diseases. Biomolecules. 2024;14(3):320,1-29. DOI: 10.3390/biom14030320.

- 100. Baker MA, Maloy WL, Zasloff M, Jacob LS. Anticancer efficacy of Magainin2 and analogue peptides. Cancer Res. 1993;53(13):3052-3057. PMID: 8319212.
- Sharma S. Melittin-induced hyperactivation of phospholipase A2 activity and calcium influx in rastransformed cells. Oncogene. 1993;8(4):939-947. PMID: 8455945.
- 102. Killion JJ, Dunn JD. Differential cytolysis of murine spleen, bone-marrow and leukemia cells by melittin reveals differences in membrane topography. Biochem Biophys Res Commun. 1986;139(1):222-227.

DOI: 10.1016/S0006-291X(86)80102-4.

 Sui SF, Wu H, Guo Y, Chen KS. Conformational changes of melittin upon insertion into phospholipid monolayer and vesicle. J Biochem. 1994;116(3): 482-487.

DOI: 10.1093/oxfordjournals.jbchem.a124550.

- 104. Peng X, Zhou C, Hou X, Liu Y, Wang Z, Peng X, et al. Molecular characterization and bioactivity evaluation of two novel bombinin peptides from the skin secretion of Oriental fire-bellied toad, Bombina orientalis. Amino acids. 2018;50(2):241-253. DOI: 10.1007/s00726-017-2509-z.
- 105. Zhao L, Tolbert WD, Ericksen B, Zhan C, Wu X, Yuan W, *et al.* Single, double and quadruple alanine substitutions at oligomeric interfaces identify hydrophobicity as the key determinant of human neutrophil alpha defensin HNP1 function. PloS One. 2013;8(11):e78937,1-14.

DOI: 10.1371/journal.pone.0078937.

- 106. Shafiee F, Rabbani M, Jahanian-Najafabadi A. Production and evaluation of cytotoxic effects of DT386-BR2 fusion protein as a novel anti-cancer agent. J Microbiol Methods. 2016;130:100-105. DOI: 10.1016/j.mimet.2016.09.004.
- 107. Pourhadi M, Jamalzade F, Jahanian-Najafabadi A, Shafiee F. Expression, purification, and cytotoxic evaluation of IL24-BR2 fusion protein. Res Pharm Sci. 2019;14(4):320-328. DOI: 10.4103/1735-5362.263556.
- 108. Shafiee F, Rabbani M, Jahanian-Najafabadi A. Optimization of the expression of DT386-BR2 fusion protein in *Escherichia coli* using response surface methodology. Adv Biomed Sci. 2017;6:22,1-6. DOI: 10.4103/2277-9175.201334.
- 109. Chen J, Xu XM, Underhill CB, Yang S, Wang L, Chen Y, *et al.* Tachyplesin activates the classic complement pathway to kill tumor cells. Cancer Res. 2005;65(11):4614-4622.

DOI: 10.1158/0008-5472.CAN-04-2253.

- 110. Ghasemi A, Ghavimi R, Momenzadeh N, Hajian S, Mohammadi M. Characterization of antitumor activity of a synthetic moronecidin-like peptide computationally predicted from the tiger tail seahorse hippocampus comes in tumor-bearing mice. Int J Pept Res Ther. 2021;27(4):2391-2401. DOI: 10.1007/s10989-021-10260-6.
- 111. Mohammadi M, Taheri B, Momenzadeh N, Salarinia R, Nabipour I, Farshadzadeh Z, *et al.* Identification and characterization of novel antimicrobial peptide from hippocampus comes by *in*

silico and experimental studies. Marine Biotechnol. 2018;20(6):718-728. DOI: 10.1007/s10126-018-9843-3.

- 112. Giuliani A, Pirri G, Nicoletto S. Antimicrobial peptides: an overview of a promising class of
- therapeutics. Cent Eur J Biol. 2007;2(1):1-33.
 DOI: 10.2478/s11535-007-0010-5.
 113. Zhang C, Yang M, Ericsson AC. Antimicrobial
- peptides: potential application in liver cancer. Front Microbiol. 2019;10:1257,1-8. DOI: 10.3389/fmicb.2019.01257.
- 114. Leite ML, da Cunha NB, Costa FF. Antimicrobial peptides, nanotechnology, and natural metabolites as novel approaches for cancer treatment. Pharmacol Ther. 2018;183:160-176.

DOI: 10.1016/j.pharmthera.2017.10.010.

115. Lv S, Sylvestre M, Prossnitz AN, Yang LF, Pun SH. Design of polymeric carriers for intracellular peptide delivery in oncology applications. Chem Rev. 2021;121(18):11653-11698.

DOI: 10.1021/acs.chemrev.0c00963.

116. Ehrenstein G, Lecar H. Electrically gated ionic channels in lipid bilayers. Q Rev Biophys. 1977;10(1):1-34.

DOI: 10.1017/S0033583500000123.

- 117. Parchebafi A, Tamanaee F, Ehteram H, Ahmad E, Nikzad H, Haddad Kashani H. The dual interaction of antimicrobial peptides on bacteria and cancer cells; mechanism of action and therapeutic strategies of nanostructures. Microb Cell Fact. 2022;21:118,1-18. DOI: 10.1186/s12934-022-01848-8.
- 118. Pouny Y, Rapaport D, Mor A, Nicolas P, Shai Y. Interaction of antimicrobial dermaseptin and its fluorescently labeled analogs with phospholipid membranes. Biochemistry. 1992;31(49):12416-12423.

DOI: 10.1021/bi00164a017.

- 119. Hilchie A, Hoskin D, Power Coombs M. Anticancer activities of natural and synthetic peptides. Adv Exp Med Biol. 2019;1117:131-147. DOI: 10.1007/978-981-13-3588-4 9.
- 120. Xiao X, Wu ZC, Chou KC. A multi-label classifier for predicting the subcellular localization of gram-negative bacterial proteins with both single and multiple sites. PloS One. 2011;6(6):e20592,1-10. DOI: 10.1371/journal.pone.0020592.
- 121. Diao Y, Han W, Zhao H, Zhu S, Liu X, Feng X, et al. Designed synthetic analogs of the α -helical peptide temporin-La with improved antitumor efficacies via charge modification and incorporation of the integrin $\alpha\nu\beta3$ homing domain. J Pept Sci. 2012;18(7):476-486. DOI: 10.1002/psc.2420.
- 122. van Hinsbergh VW, Collen A, Koolwijk P. Angiogenesis and anti-angiogenesis: perspectives for the treatment of solid tumors. Ann Oncol. 1999;10:4,60-63. PMID: 10436787.

123. Gacche RN, Meshram RJ. Targeting tumor micro-

environment for design and development of novel anti-angiogenic agents arresting tumor growth. Prog Biophys Mol Biol. 2013;113(2):333-354. DOI: 10.1016/j.pbiomolbio.2013.10.001. 124. Saharinen P, Eklund L, Pulkki K, Bono P, Alitalo K. VEGF and angiopoietin signaling in tumor angiogenesis and metastasis. Trends Mol Med. 2011;17(7):347-362.

DOI: 10.1016/j.molmed.2011.01.015.

- 125. Yi ZF, Cho SG, Zhao H, Wu Yy, Luo J, Li D, *et al*. A novel peptide from human apolipoprotein (a) inhibits angiogenesis and tumor growth by targeting c-Src phosphorylation in VEGF-induced human umbilical endothelial cells. Int J Cancer. 2009;124(4):843-852. DOI: 10.1002/ijc.24027.
- 126. Jang JP, Jung HJ, Han JM, Jung N, Kim Y, Kwon HJ, et al. Two cyclic hexapeptides from *Penicillium* sp. FN070315 with antiangiogenic activities. PLoS One. 2017;12(9):e0184339,1-15. DOI: 10.1371/journal.pone.0184339.
- 127. Zhang Y, Nicolau A, Lima CF, Rodrigues LR. Bovine lactoferrin induces cell cycle arrest and inhibits mTOR signaling in breast cancer cells. Nutr Cancer. 2014;66(8):1371-1385. DOI: 10.1090/01605581.2014.056260

DOI: 10.1080/01635581.2014.956260.

128. Wolf JS, Li G, Varadhachary A, Petrak K, Schneyer M, Li D, *et al.* Oral lactoferrin results in T cell-dependent tumor inhibition of head and neck squamous cell carcinoma *in vivo*. Clin Cancer Res. 2007;13(5):1601-1610.

DOI: 10.1158/1078-0432.CCR-06-2008.

129. Li X, Meng Y, Plotnikoff NP, Youkilis G, Griffin N, Wang E, *et al.* Methionine enkephalin (MENK) inhibits tumor growth through regulating CD4+ Foxp3+ regulatory T cells (Tregs) in mice. Cancer Biol Ther. 2015;16(3):450-459.

DOI: 10.1080/15384047.2014.1003006.

 Zhao D, Plotnikoff N, Griffin N, Song T, Shan F. Methionine enkephalin, its role in immunoregulation and cancer therapy. Int Immunopharmacol. 2016;37:59-64.

DOI: 10.1016/j.intimp.2016.02.015.

- 131. Zagon IS, Donahue RN, McLaughlin PJ. Opioid growth factor-opioid growth factor receptor axis is a physiological determinant of cell proliferation in diverse human cancers. Am J Physiol Regul Integr Comp Physiol. 2009;297(4):R1154-R1161. DOI: 10.1152/ajpregu.00414.2009.
- 132. Huang Y, Feng Q, Yan Q, Hao X, Chen Y. Alphahelical cationic anticancer peptides: a promising candidate for novel anticancer drugs. Mini Rev Med Chem. 2015;15(1):73-81.

DOI: 10.2174/1389557514666141107120954.

- 133. Li H, Kolluri SK, Gu J, Dawson MI, Cao X, Hobbs PD, *et al.* Cytochrome c release and apoptosis induced by mitochondrial targeting of nuclear orphan receptor TR3. Science. 2000;289(5482):1159-1164. DOI: 10.1126/science.289.5482.1159.
- 134. Ekundayo BE, Obafemi TO, Adewale OB, Obafemi BA, Oyinloye BE, Ekundayo SK. Oxidative stress, endoplasmic reticulum stress and apoptosis in the pathology of Alzheimer's disease. Cell Biochem Biophys. 2024;82(2):457-477. DOI: 10.1007/s12013-024-01248-2.
- 135. Liu M, Zhao X, Zhao J, Xiao L, Liu H, Wang C, *et al.* Induction of apoptosis, G0/G1 phase arrest and

microtubule disassembly in K562 leukemia cells by Mere15, a novel polypeptide from *Meretrix meretrix* Linnaeus. Mar Drugs. 2012;10(11):2596-2607. DOI: 10.3390/md10112596.

- 136. Huang TC, Lee JF, Chen JY. Pardaxin, an antimicrobial peptide, triggers caspase-dependent and ROS-mediated apoptosis in HT-1080 cells. Mar Drugs. 2011;9(10):1995-2009. DOI: 10.3390/md9101995.
- 137. Qu B, Yuan J, Liu X, Zhang S, Ma X, Lu L. Anticancer activities of natural antimicrobial peptides from animals. Front Microbiol. 2024;14:1321386,1-17.

DOI: 10.3389/fmicb.2023.1321386.

138. Harris F, Dennison SR, Singh J, Phoenix DA. On the selectivity and efficacy of defense peptides with respect to cancer cells. Med Res Rev. 2013;33(1):190-234.

DOI: 10.1002/med.20252.

 Schweizer F. Cationic amphiphilic peptides with cancer-selective toxicity. Eur J Pharmacol. 2009;625(1-3):190-194.
 DOL: 10.1016/i.cipher.2000.08.042

DOI: 10.1016/j.ejphar.2009.08.043.

140. Dobrzyńska I, Szachowicz-Petelska B, Sulkowski S, Figaszewski Z. Changes in electric charge and phospholipids composition in human colorectal cancer cells. Mol Cell Biochem. 2005;276(1-2):113-119.

DOI: 10.1007/s11010-005-3557-3.

141. Lee E, Rosca EV, Pandey NB, Popel AS. Small peptides derived from somatotropin domaincontaining proteins inhibit blood and lymphatic endothelial cell proliferation, migration, adhesion and tube formation. Int J Biochem Cell Biol. 2011;43(12):1812-1821.

DOI: 10.1016/j.biocel.2011.08.020.

142. Mader JS, Hoskin DW. Cationic antimicrobial peptides as novel cytotoxic agents for cancer treatment. Expert Opin Investig Drugs. 2006;15(8):933-946. DOI: 10.1517/13543784.15.8.933.

143. Mulukutla A, Shreshtha R, Deb VK, Chatterjee P, Jain U, Chauhan N. Recent advances in antimicrobial peptide-based therapy. Bioorg Chem. 2024;145:107151.

DOI: 10.1016/j.bioorg.2024.107151.

144. Jiang R, Du X, Lönnerdal B. Comparison of bioactivities of talactoferrin and lactoferrins from human and bovine milk. J Pediatr Gastroenterol Nutr. 2014;59(5):642-652.

DOI: 10.1097/MPG.00000000000481.

145. Roudi R, Syn NL, Roudbary M. Antimicrobial peptides as biologic and immunotherapeutic agents against cancer: a comprehensive overview. Front immunol. 2017;8:1320,1-10.

DOI: 10.3389/fimmu.2017.01320.

- 146. Tyagi A, Tuknait A, Anand P, Gupta S, Sharma M, Mathur D, *et al.* CancerPPD: a database of anticancer peptides and proteins. Nucleic Acids Res. 2015;43(D1):D837-D843. DOI: 10.1093/nar/gku892.
- 147. Hajisharifi Z, Piryaiee M, Beigi MM, Behbahani M, Mohabatkar H. Predicting anticancer peptides

with Chou's pseudo amino acid composition and investigating their mutagenicity *via* Ames test. J Theor Biol. 2014;341:34-40.

DOI: 10.1016/j.jtbi.2013.08.037.

- 148. Grisoni F, Neuhaus CS, Gabernet G, Müller AT, Hiss JA, Schneider G. Designing anticancer peptides by constructive machine learning. ChemMedChem. 2018;13(13):1300-1302. DOI: 10.1002/cmdc.201800204.
- 149. Kozłowska K, Nowak J, Kwiatkowski B, Cichorek M. ESR study of plasmatic membrane of the transplantable melanoma cells in relation to their biological properties. Exp Toxicol Pathol. 1999;51(1):89-92.

DOI: 10.1016/S0940-2993(99)80074-8.

150. Zhang W, Li J, Liu LW, Wang KR, Song JJ, Yan JX, *et al.* A novel analog of antimicrobial peptide Polybia-MPI, with thioamide bond substitution, exhibits increased therapeutic efficacy against cancer and diminished toxicity in mice. Peptides. 2010;31(10):1832-1388.

DOI: 10.1016/j.peptides.2010.06.019.

- 151. Gaspar D, Veiga AS, Sinthuvanich C, Schneider JP, Castanho MA. Anticancer peptide SVS-1: efficacy precedes membrane neutralization. Biochemistry. 2012;51(32):6263-6265. DOI: 10.1021/bi300836r.
- 152. Yang QZ, Wang C, Lang L, Zhou Y, Wang H, Shang DJ. Design of potent, non-toxic anticancer peptides based on the structure of the antimicrobial peptide, temporin-1CEa. Arch Pharm Res. 2013;36(11):1302-1310.

DOI: 10.1007/s12272-013-0112-8.

153. Lemeshko VV. Electrical potentiation of the membrane permeabilization by new peptides with anticancer properties. Biochim Biophys Acta. 2013;1828(3):1047-1056.

DOI: 10.1016/j.bbamem.2012.12.012.

- 154. Hu C, Chen X, Zhao W, Chen Y, Huang Y. Design and modification of anticancer peptides. Drug Des. 2016;5(3):1000138,1-10. DOI: 10.4172/2169-0138.1000138.
- 155. Boohaker RJ, Lee MW, Vishnubhotla P, Perez JM, Khaled AR. The use of therapeutic peptides to target and to kill cancer cells. Curr Med Chem. 2012;19(22):3794-3804.

DOI: 10.2174/092986712801661004.

156. Bidwell GL 3rd, Raucher D. Therapeutic peptides for cancer therapy. Part I- peptide inhibitors of signal transduction cascades. Expert Opin Drug Deliv. 2009;6(10):1033-1047.

DOI: 10.1517/17425240903143745.

- 157. Huang Y, Li X, Sha H, Zhang L, Bian X, Han X, *et al.* Tumor-penetrating peptide fused to a proapoptotic peptide facilitates effective gastric cancer therapy. Oncol Rep. 2017;37(4):2063-2070. DOI: 10.3892/or.2017.5440.
- Duvic M, Talpur R. Optimizing denileukin diftitox (Ontak) therapy. 2008;4(4):457-469. DOI: 10.2217/14796694.4.457
- 159. Ghavimi R, Akbari V, Jahanian-Najafabadi A. Production and evaluation of *in vitro* and *in vivo* effects of P28-IL24, a promising anti-breast cancer

fusion protein. Int J Pept Res Ther. 2021;27(4): 2583-2594.

DOI: 10.1007/s10989-021-10275-z.

- 160. Liu M, Wang H, Liu L, Wang B, Sun G. Melittin-MIL-2 fusion protein as a candidate for cancer immunotherapy. J Transl Med. 2016;14(1):1-12. DOI: 10.1186/s12967-016-0910-0.
- 161. Aguirre TA, Teijeiro-Osorio D, Rosa M, Coulter I, Alonso M, Brayden D. Current status of selected oral peptide technologies in advanced preclinical development and in clinical trials. Adv Drug Deliv Rev. 2016;106(Pt B):223-241. DOI: 10.1016/j.addr.2016.02.004.
- 162. Wang L, Wang N, Zhang W, Cheng X, Yan Z, Shao G, *et al.* Therapeutic peptides: current applications and future directions. Signal Transduct Target Ther. 2022;7(1):48,1-27.
- DOI: 10.1038/s41392-022-00904-4.
- 163. Baguley BC. Multiple drug resistance mechanisms in cancer. Mol Biotechnol. 2010;46(3):308-316. DOI: 10.1007/s12033-010-9321-2.
- 164. Kakde D, Jain D, Shrivastava V, Kakde R, Patil A. Cancer therapeutics-opportunities, challenges and advances in drug delivery. J Appl Pharm Sci. 2011;1(9):1-10.

DOI: 10.4155/tde-2020-0079.

165. Gordon YJ, Romanowski EG, McDermott AM. A review of antimicrobial peptides and their therapeutic potential as anti-infective drugs. Curr Eye Res. 2005;30(7):505-515.

DOI: 10.1080/02713680590968637.

166. Kao C, Lin X, Yi G, Zhang Y, Rowe-Magnus DA, Bush K. Cathelicidin antimicrobial peptides with reduced activation of Toll-like receptor signaling have potent bactericidal activity against colistinresistant bacteria. mBio. 2016;7(5):e01418e01416,1-10.

DOI: 10.1128/mbio.01418-16.

- 167. Ghadiri N, Javidan M, Taştan Ö, Liao Z, Ganjalıkhani-Hakemi M. Bioactive peptides: an alternative therapeutic approach for cancer management. Front Immunol.2024;15:1310443,1-18. DOI: 10.3389/fimmu.2024.1310443.
- Renukuntla J, Vadlapudi AD, Patel A, Boddu SH, Mitra AK. Approaches for enhancing oral bioavailability of peptides and proteins. Int J Pharm. 2013;447(1-2):75-93.

DOI: 10.1016/j.ijpharm.2013.02.030.

 Ismail R, Csoka I. Novel strategies in the oral delivery of antidiabetic peptide drugs–Insulin, GLP 1 and its analogs. Eur J Pharm Biopharm. 2017;115:257-267.

DOI: 10.1016/j.ejpb.2017.03.015.

- 170. Uggerhøj LE, Poulsen TJ, Munk JK, Fredborg M, Sondergaard TE, Frimodt-Moller N, *et al.* Rational design of alpha-helical antimicrobial peptides: do's and don'ts. ChemBioChem. 2015;16(2):242-253. DOI: 10.1002/cbic.201402581.
- 171. Kelly GJ, Kia AFA, Hassan F, O'Grady S, Morgan MP, Creaven B, *et al.* Polymeric prodrug combination to exploit the therapeutic potential of antimicrobial peptides against cancer cells. Org Biomol Chem. 2016;14(39):9278-9286.

- 172. Dąbrowska K, Kaźmierczak Z, Majewska J, Miernikiewicz P, Piotrowicz A, Wietrzyk J, *et al.* Bacteriophages displaying anticancer peptides in combined antibacterial and anticancer treatment. Future Microbiol. 2014;9(7):861-869. DOI: 10.2217/fmb.14.50.
- 173. Hao X, Yan Q, Zhao J, Wang W, Huang Y, Chen Y. TAT modification of alpha-helical anticancer peptides to improve specificity and efficacy. PLoS One. 2015;10(9):e0138911,1-13. DOI: 10.1371/journal.pone.0138911.
- 174. Darwish W. Polymers for enhanced photodynamic cancer therapy: phthalocyanines as a photosensitzer model. Polym Adv Technol. 2021;32(3):919-930. DOI: 10.1002/pat.5154.
- 175. Kang HK, Kim C, Seo CH, Park Y. The therapeutic applications of antimicrobial peptides (AMPs): a patent review. J Microbiol. 2017; 55(1):1-12.

DOI: 10.1007/s12275-017-6452-1.

- 176. Trinidad-Calderón PA, Varela-Chinchilla CD, García-Lara S. Natural peptides inducing cancer cell death: Mechanisms and properties of specific candidates for cancer therapeutics. Molecules. 2021;26(24):7453,1-21.
 - DOI: 10.3390/molecules26247453.
- 177. Vitale I, Yamazaki T, Wennerberg E, Sveinbjørnsson B, Rekdal Ø, Demaria S, *et al.* Targeting cancer heterogeneity with immune responses driven by oncolytic peptides. Trends Cancer. 2021;7(6):557-572.

DOI: 10.1016/j.trecan.2020.12.012.

- 178. Darabi F, Saidijam M, Nouri F, Mahjub R, Soleimani M. Anti-CD44 and EGFR dual-targeted solid lipid nanoparticles for delivery of doxorubicin to triple-negative breast cancer cell line: preparation, statistical optimization, and *in vitro* characterization. Biomed Res Int. 2022;2022(1):6253978,1-13. DOI: 10.1155/2022/6253978.
- 179. Faraji N, Arab SS, Doustmohammadi A, Daly NL, Khosroushahi AY. ApInAPDB: a database of apoptosis-inducing anticancer peptides. Sci Rep. 2022;12(1):21341,1-7.

DOI: 10.1038/s41598-022-25530-6.

180. Ghavimi R, Mohammadi E, Akbari V, Shafiee F, Jahanian-Najafabadi A. *In silico* design of two novel fusion proteins, p28-IL-24 and p28-M4, targeted to breast cancer cells. Res Pharm Sci. 2020;15(2): 200-208.

DOI: 10.4103/1735-5362.283820.

181. Chen Z, Wang R, Guo J, Wang X. The role and future prospects of artificial intelligence algorithms in peptide drug development. Biomed Pharmacother. 2024;175:116709,1-13.

DOI: 10.1016/j.biopha.2024.116709.

182. Aguilera-Puga MdC, Cancelarich NL, Marani MM, de la Fuente-Nunez C, Plisson F. Accelerating the discovery and design of antimicrobial peptides with artificial intelligence. Method Mol Biol. 2024;2714:329-352.

DOI: 10.1007/978-1-0716-3441-7_18.

- Anti-cancer peptides as cancer therapeutics
- 183. Ghaly G, Tallima H, Dabbish E, Badr ElDin N, Abd El-Rahman MK, Ibrahim MA, *et al.* Anti-cancer peptides: status and future prospects. Molecules. 2023;28(3):1148,1-27.

DOI: 10.3390/molecules28031148.

184. Huang KY, Tseng YJ, Kao HJ, Chen CH, Yang HH, Weng SL. Identification of subtypes of anticancer peptides based on sequential features and physicochemical properties. Sci Rep. 2021; 11(1):1-13.

DOI: 10.1038/s41598-021-93124-9.

185. Fu B, Zhang Y, Long W, Zhang A, Zhang Y, An Y, *et al.* Identification and characterization of a novel phage display-derived peptide with affinity for human brain metastatic breast cancer. Biotechnol Lett. 2014;36(11):2291-2301.
DOI: 10.1007/c10520.014.1008.0

DOI: 10.1007/s10529-014-1608-0.

- 186. Bakare OO, Gokul A, Wu R, Niekerk LA, Klein A, Keyster M. Biomedical relevance of novel anticancer peptides in the sensitive treatment of cancer. Biomolecules. 2021;11(8):1120,1-17. DOI: 10.3390/biom11081120.
- 187. Lath A, Santal AR, Kaur N, Kumari P, Singh NP. Anti-cancer peptides: their current trends in the development of peptide-based therapy and anti-tumor drugs. Biotechnol Genet Eng Rev. 2023;39(1):45-84. DOI: 10.1080/02648725.2022.2082157.
- 188. Conibear AC, Schmid A, Kamalov M, Becker CF, Bello C. Recent advances in peptide-based approaches for cancer treatment. Curr Med Chem. 2020;27(8):1174-1205.

DOI: 10.2174/0929867325666171123204851.

 Rodríguez FD, Coveñas R. Antitumor strategies targeting peptidergic systems. Encyclopedia. 2024;4(1):478-487.

DOI: 10.3390/encyclopedia4010031.

190. Nhàn NTT, Yamada T, Yamada KH. Peptidebased agents for cancer treatment: current applications and future directions. Int J Mol Sci. 2023;24(16):12931,1-35.

DOI: 10.3390/ijms241612931.

- 191. Ruggirello C, Mörl K, Beck-Sickinger AG. Peptides for therapeutic applications-challenges and chances. Pure Appl Chem. 2024;96(1):91-103. DOI: 10.1515/pac-2024-0104.
- 192. Ekinci M, Magne TM, Alencar LMR, Fechine PBA, Santos-Oliveira R, Ilem-Özdemir D. Molecular imaging for lung cancer: exploring small molecules, peptides, and beyond in radiolabeled diagnostics. Pharmaceutics. 2024;16(3):404,1-21. DOI: 10.3390/pharmaceutics16030404.
- 193. Khalily MP, Soydan M. Peptide-based diagnostic and therapeutic agents: where we are and where we are heading? Chem Biol Drug Des. 2023;101(3): 772-793.

DOI: 10.1111/cbdd.14180.

194. Noei A, Nili-Ahmadabadi A, Soleimani M. The enhanced cytotoxic effects of the p28-apoptin chimeric protein as a novel anti-cancer agent on breast cancer cell lines. Drug Res. 2019;69(03): 144-150.

DOI: 10.1055/a-0654-4952.