

Antispasmodic activity of isovanillin and isoacetovanillon in comparison with *Pycnocycla spinosa* Decne.exBoiss extract on rat ileum

H. Sadraei^{1,*}, M. Ghanadian², G. Asghari² and E. Madadi^{1,2}

¹Department of Pharmacology & Toxicology and Isfahan Pharmaceutical Sciences Research Center, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, I.R. Iran. ²Department of Pharmacognosy and Isfahan Pharmaceutical Sciences Research Center, Isfahan University of Medical Sciences, Isfahan, I.R. Iran.

Abstract

Isovanillin and isoacetovanillon are two components found in *P. spinosa* Decne.exBoiss extract with no previously reported effect on ileum contractions. Spasmolytic effect of isovanillin and isoacetovanillon were examined on response to electrical field stimulation (EFS), acetylcholine (ACh) and 5-HT in strips of rat ileum. Longitudinal ileum strips were set up in an organ bath containing oxygenated Tyrode's solution. All strips that was contracted in response to EFS, acetylcholine or 5-HT showed relaxation in the presence of isovanillin (5-320 μ g/ml), or isoacetovanillon (5-320 μ g/ml). Isovanillin and isoacetovanillon inhibited the response to 5-HT with IC₅₀ values of 356±50 μ M and 622±110 μ M respectively. They reduced the response to EFS without significantly affecting the acetylcholine response. *P. spinosa* extract (5-160 μ g/ml) in a concentration dependent manner reduced the response to 5-HT, acetylcholine and EFS. This study demonstrated that isovanillin and isoacetovanillon are relaxant of ileum contractions induced by 5-HT and EFS and they have contribution to the relaxant effect of *P. spinosa* extract but other components are responsible for the inhibition of acetylcholine by the extract.

Keywords: Isovanillin; Isoacetovanillon; Pycnocycla spinosa extract; Propantheline; Acetylcholine; 5-HT

INTRODUCTION

Pycnocycla spinosa Decne.exBoiss is a wild plant which grows in Middle East including Iran (1). Hydroalcoholic extract of *P. spinosa* has both antispasmodic and anti-diarrhoeal effect (2-5). The extract of *P. spinosa* contains many active substances including flavonoids, glycosides, tannins and alkaloids–like substances (3,4). Fractionations of extracts also indicated presence of a number of active substances in the extract (6).

Further separation and isolation lead to identification of active substances in the extract including isovanillin and isoacetovanillon (Fig. 1) which may be responsible for antispasmodic effect of *P. spinosa* extract (7).

Although, these two substances are already known as plant material constituents, however,

their pharmacological effect on ileum motility has not been reported, so far. Therefore, the aim of this research was to investigate the antispasmodic of isovanillin and isoacetovanillon in comparison with *P*. *spinosa* extract and a reference drug; propantheline.



Fig. 1. Chemical structure of isoacetovanillon (A) and isovanillin (B).

*Corresponding author: H. Sadraei, this paper is extracted from the Pharm.D thesis No. 190104 Tel. 0098 311 792 2608, Fax. 0098 311 6680011 Email: sadraei@pharm.mui.ac.ir

MATERIALS AND METHODS

Drugs and solutions

Drugs used were as follows: Acetylcholine, 5-hydroxytriptamine (5-HT),and propantheline (Sigma, Germany), lidocaine (Pasteur, Tehran) isovanillin and isoacetovanillon (Sigma, China).

Acetylcholine (250 μ M), 5-HT (100 μ M), lidocaine (7.4 mM) and propantheline (1 mM) stock solutions were made up in distilled water. Isovanillin and isoacetovanillon were made up as 20 mg/ml stock solution in dimethyl sulphoxide (DMSO). Further serial dilutions were prepared in distilled water. Tyrode's solution with the following composition: NaCl; 136.9, KCl; 2.68, CaCl₂;1.8, MgCl₂; 1.05, NaHCO₃; 11.9, NaH₂PO₄; 0.42 and glucose 5.55 mM were made up in distilled water. All the chemicals were from Merck (Germany) unless stated otherwise.

Plant extract

The aerial parts of Pycnocycla spinosa Decne.exBoiss. var. spinosa (Fam. Umbelliferae) were collected from base of Sofah mountain in Isfahan, Iran. The plant was identified by Dr Mehregan, plant taxonomist (Tehran Azad University). Α voucher specimen (A24) of the plant was deposited in the herbarium of the School of Pharmacy and Pharmaceutical sciences at the Isfahan University of Medical Sciences, Iran.

Studies on ileum

Male Wistar rats (200-220 g) were killed and longitudinal strips of ileum were taken and placed in oxygenated Tyrode's solution. Each strip was suspended between parallel platinum electrodes in 50 ml organ bath. The bathing fluid was Tyrode's solution at 37°C and gassed with oxygen. The ileum tension was monitored with an isotonic transducer and displayed on a pen recorder (Harvard) device. Each preparation was subjected to a constant 1g weight tension.

Electrical field stimulation (EFS) was performed using rectangular pulses produced by a stimulator (Designed in Isfahan). Tissues were stimulated with maximum of 6 volts using 1s trains of stimuli at 50 Hz. Acetylcholine (0.5 μ M) and 5-HT (2 μ M) were added into bath with contact time of 20 s. Stimulation was performed once every 10 or 15 min as appropriate.

After the tissue baseline and responses were stabilized, pure compounds or extract were directly added into the bath using two fold increments in concentration unless stated. Value in the text shows the final bath concentration. Adequate time was permitted after adding each agent for equilibration of the response before further testing.

Analysis of data

Contraction to EFS or to added spasmogens were measured relative to the tissue baseline and expressed as percentage of initial response prior to addition of testing agent. The IC_{50} value (drug concentration causing 50% of maximum response) was determined by plotting a full concentration response curve for each tissue. Potency of the compounds was expressed as pD_2 value (the negative log_{10} of the molar IC_{50}). Values are presented as mean \pm standard error of mean (SEM). Statistical analysis was performed using Student's t-test and/or one way analysis of variance (ANOVA) as appropriate. Sigma Plot (version 11) computer program was used for statistical analysis and plotting of graphs.

RESULTS

The response of ileum strip to electrical field stimulation was a rapid contraction (EFS-1), followed by relaxation and 30-90 s later a second contraction (EFS-2) which peaks up and returned to baseline level relatively slowly. Acetylcholine and 5-HT caused a rapid contraction the amplitude of which slightly falls off before it was washout. The response to EFS was relatively smaller than the response to acetylcholine or 5-HT. The responses to EFS were reduced by 5 and 50 µM lidocaine while the acetylcholine and 5-HT responses were not significantly affected. Larger responses to electrical stimulation could be obtained by using a larger pulse, but such response may not be abolished by lidocaine. At 500 µM bath concentration lidocaine however inhibited the response to acetylcholine and 5-HT showing a direct effect of lidocaine on smooth muscle.



Fig. 2. Inhibitory effect of propantheline on tension development in the isolated ileum of rat treated with 5-HT (2 µM), acetylcholine (ACh, 500 nM) and electrical field stimulation (EFS: 6 V, 50 Hz, 1 s duration). Each data point is mean \pm SEM (n=6). Ordinant scales: spasm remaining as a % of the contraction prior to propantheline addition. Abscissa scales: \log_{10} concentration of propantheline. Each point is mean and the vertical lines show the SEM. EFS-1=initial contractile response. EFS-2=secondary contractile response. Stars show significant differences at corresponding propantheline concentration with 5-HT. *P<0.05, **P<0.01, ***P<0.001 (Student's t-test).



Fig. 4. Inhibitory effect of isovanillin on tension development in the isolated ileum of rat treated with 5-HT (2 μ M), acetylcholine (ACh, 500 nM) and electrical field stimulation (EFS: 6 V, 50 Hz, 1 s duration). Each data point is mean \pm SEM (n=6). Ordinant scales: spasm remaining as a % of the contraction prior to isovanillin addition. Abscissa scales: log₁₀ concentration of isovanillin. Each point is mean and the vertical lines show the SEM. EFS-1= initial contractile response. EFS-2= secondary contractile response. The variation in ACh response is not statistically significant but inhibition of 5-HT and EFS responses are significant (P<0.001, ANOVA).



Fig. 3. Log concentration inhibitory response curve of *P. spinosa* extract on tension development in the isolated ileum of rat treated with 5-HT (2 μ M), acetylcholine (ACh, 500 nM) and electrical field stimulation (EFS: 6 V, 50 Hz, 1 s duration). Each data point is mean \pm SEM (n=6). Ordinant scales: spasm remaining as a % of the contraction prior to extract addition. Abscissa scales: \log_{10} concentration of *P. spinosa* extract. Each point is mean and the vertical lines show the SEM. EFS-1=initial contractile response. EFS-2=secondary contractile response. There was no statistically significant changes in vehicle treated time matched controls (curves not shown) but the reduction of responses by extract is significant (P<0.001, ANOVA).



Fig. 5. Inhibitory effect of isoacetovanillon on tension development in the isolated ileum of rat treated with 5-HT (2 μ M), acetylcholine (ACh, 500 nM) and electrical field stimulation (EFS: 6 V, 50 Hz, 1 s duration). Each data point is mean \pm SEM (n=6). Ordinant scales: spasm remaining as a % of the contraction prior to isoacetovanillon addition. Abscissa scales: \log_{10} concentration of isoacetovanillon. Each point is mean and the vertical lines show the SEM. EFS-1= initial contractile response. EFS-2= secondary contractile response. The variation in ACh response is not statistically significant but inhibition of 5-HT and EFS responses are significant (P<0.001, ANOVA).

All the tested tissues have shown repeatable contractile response to EFS, acetylcholine and/or 5-HT. Tissues with no consistent response were not further pursued.

Propantheline at concentration which totally blocked the response to acetylcholine (0.5 nM-32 nM) partially reduced the contraction in response to EFS without affecting the response to 5-HT (Fig. 2). The remaining response to EFS shows the presence of non-adrenergic non-cholinergic (NANC) responses. There was no significant changes in the contractile response in vehicle treated time matched control tissues.

P. spinosa extract (10-160 µg/ml) concentration dependently inhibited the response to 5-HT (IC₅₀=28 ± 2 µg/ml, n=6), acetylcholine (IC₅₀=55 ± 8 µg/ml, n=6) and EFS (EFS-1, IC₅₀=27 ± 4 µg/ml, n=6), at its highest used bath concentration (320 µg/ml), the extract abolished the response to above stimuli (Fig. 3).

Isovanillin (5-320 µg/ml) and isoacetovanillon (5-320 µg/ml) concentration dependently inhibited the contractile responses to 5-HT with IC₅₀ values of $54 \pm 8 \mu \text{g/ml}$ (pD₂=3.5 \pm 0.07, n=6) and 103 \pm 19 µg/ml (pD₂=3.3 \pm 0.1, n=6), respectively (see Figs. 4, 5). Both isovanillin and isoacetovanillon reduced the responses to EFS but the inhibitory effect was not complete (Figs. 4, 5). Isovanillin and isoacetovanillon at concentra-tions which inhibited the response to 5-HT did not affected the contractile response to acetylcholine (Figs. 4, 5). In the vehicle treated time matched control tissues, DMSO (0.03%-1.6%) only caused a slight increase in contractile responses to acetylcholine, 5-HT and EFS.

DISCUSSION

Studies have shown that, a seris of phenols that are structurally related to 3-*t*-butyl-4hydroxy-anisole (BHA), have antispasmodic and spasmolytic activity in rat ileum longitudinal muscle (8). Preliminary data indicate that BHA is an active substance which also has antispasmodic effects on vascular smooth muscle (9,10). Isovanillin is a phenolic aldehyde (Fig. 1), an organic compound and isomer of vanillin and it is found in a number

plants including Pimpinella of anisum (apiaceae), Evodia rutaecarpa (apiaceae) and (apiaceae) (11-13). *Coriandrum* sativum Isovanillin is a selective inhibitor of aldehvde oxidase (14). It is not a substrate of that enzyme, and is metabolized by aldehyde dehydrogenase into isovanillic acid (14). Despite presence of isovanillin in number of plant material, however so far there isn't any official report on its pharmacological action on gastrointestinal function. Isoacetovanillon (Fig. 1) is also found in plant materials (15) but its pharmacological activity is not documented except a short report about analgesic effect and inhibitory action on the gastrointestinal motility as well as low degree of antibacterial activity against Escherichia coli and Shigella flexneri (15).

An isolated organ bath assay was used as pharmacological screening tool to assess concentration-response relationships of isovanillin and isoacetovanillon in rat ileum. These two compounds have been identified in extract of P. spinosa (7). The complex responses of the organ bath assay can be studied while controlling several physiological parameters. Therefore, effects of compounds with unknown molecular targets can be studied because all important tissue-specific expression and modification of molecular targets remain intact in organ bath preparation. Isovanillin, isoacetovanillon and the hydroalcoholic extract of P. spinosa concentration dependently induced inhibited the contraction bv acetylcholine, 5-HT and EFS. Two former induces contractions via activating muscarinic M₃ receptors and serotonergic 5-HT₂ receptors repectively on ileum smooth muscle cells, while EFS stimulate the mesenteric neurons within the tissue including NANC nerves which have been described in all regions of the gastrointestinal tract (16-20).

Propantheline which is a muscarinic receptor antagonist at concentrations which totally removed the response to acetylcholine, only partially reduced the contractile response to EFS. As it would be expected, propantheline had no effect on 5-HT response. On the other hand. isovanillin and isoacetovanillon totally removed the ileum contractile responses to EFS, indicating that it also can remove contraction induced by

neurotransmitters released from NANC nerves. In addition, the contractile response to 5-HT was also removed. Furthermore, isovanillin and isoacetovanillon are relaxant of rat ileum contraction induced by KCl (7) while propantheline and atropine have no effect on contraction induced by KCl (21,22). When these results are compared, it could be concluded that the mechanism of action of isovanillin and isoacetovanillon is somehow different from that of simple muscarinic antagonism.

Since isovanillin and isoacetovanillon both inhibited the response to different spasmogens, a more general intracellular mechanism might be involved because these compounds inhibited contraction due to release of Ca²⁺ ions from intracellular store (acetylcholine and 5-HT) or due to activation of voltage gated calcium channels (KCl) (7). Isoacetovanillon is a structural isomer of acetovanillon which has been widely used as an NADPH oxidase inhibitor in many experimental models (23-26). Isovanillin is also an inhibitor of aldehyde oxidase in the liver slice (14). Since isovanillin and isoacetovanillon have many structural similarities, it is possible that these two compounds act on similar intracellular target or membrane receptors, probably inhibiting specific enzyme involved in smooth muscle contraction. However, the exact mechanism of action needs to be investigated.

The total hydroalcoholic extract of *P*. spinosa was as potent as the isovanillin and isoacetovanillon in inhibiting rat ileum contraction. As isovanillin and isoacetovanillon only compose a small fraction of *P*. spinosa extract, it is clear that other active components exist in the extract with significant contribution in relaxant effect of *P*. spinosa extract. This can be supported by the reports that on gram weight bases some fractions of *P*. spinosa extract are about 10 times more active than isovanillin and isoacetovanillon (7).

CONCLUSION

In this study we have shown that isovanillin and isoacetovanillon are potent relaxant of rat isolated ileum. As these two components are found in *P. spinosa* extract it can be concluded that isovanillin and isoacetovanillon have significant contribution in antispasmodic action of *P. spinosa* extract and they could be used as an alternative remedy for treatment of spasmodic gastrointestinal disorders.

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REFERENCES

- 1. Jalili A, Jamzad Z. Red data book of Iran, A preliminary survey of endemic, rare and endangered plant species in Iran. Tehran: Research Institute of Forests and Rangelands; 1999. p. 689-690.
- 2. Sadraei H, Asghari G, Naddafi A. Relaxant effect of essential oil and hydro-alcoholic extract of *Pycnocycla spinosa* Decne. exBoiss. on ileum contraction. Phytother Res. 2003;17:645-649.
- Sadraei H, Asghari G, Hekmatti AA. Antispasmodic effect of three fraction of hydroalcoholic extract of *Pycnocycla spinosa*. J Ethnopharmacol. 2003;86:187-190.
- 4. Sadraei H, Asghari G, Khazael M. Relaxant effect of four fractions separated from alkaloid extract of *Pycnocycla spinosa* on rat isolated ileum. Res Pharm Sci. 2008;3:79-86.
- 5. Sadraei H, Asghari G, Shams M. Antidiarrhoeal action of hydroalcoholic extract of *Pycnocycla spinosa* in comparison with loperamide and dicyclomine. Iranian J Pharm Res. 2011;10:835-841.
- 6. Sadraei H, Asghari G, Behzad S. Bioactivity-guided isolation of spasmolytic components of *Pycnocycla spinosa*. Decne exBoiss. Res Pharm Sci. 2011;6:81-86.
- 7. Jahed M. Quantitative comparison of anti-spasmodic action of fractions separated from *Pycnocycla spinosa* extract on rat ileum using a bioassay technique. PharmD [Thesis], Isfahan University of Medical Sciences; 2013.
- 8. Sgaragli GP, Valoti M, Gorelli B. Calcium antagonist and antiperoxidant properties of some hindered phenoles. Br J Pharmacol. 1993;110:369-377.
- 9. Gorelli B, Pessina F, Fusi F. Calcium antagonist property of some hindered phenols on rat aorta rings. Pharmacol Res. 1995;31:206.
- Fusi F, Valoti M, Frosini F. 2,5-di-t-butyl-1,4benzohydroquinone BHQ induces endotheliumdependent relaxation of rat thoracic aorta. Eur J Pharmacol. 1999;366:181-187.
- 11. Birgitt K, Jurgen R. S-adenosyl-l-methionine: Anol-Omethyltransferase activity in organ cultures of *Pimpinella anisum*. Phyto Chem. 1996;42:397-403.
- Wang Q, Liang J, Chen J. Chemical constituents of Evodia rutaecarpa. J China Pharm Uni. 2005;36: p. 520.

- Singh G, Maurya S, Lampasona M. Chemical composition, antifungal, antioxidant and sprout suppressant activities of coriander (*Coriandrum sativum*) essential oil and its oleoresin. Flavour Fragr J. 2006;21:472-479.
- Panoutsopoulos GI, Beedham C. Metabolism of isovanillin by aldehyde oxidase, xanthine oxidase, aldehyde dehydrogenase and liver slices. Pharmacol. 2005;73:199-208.
- 15. Sun FZ, Cai M, Lou FC. Analegesic effect and gastrointestinal motility inhibitory action of 3-hydroxy-4methoxy-acetophenone from *Cynanchum paniculatum* (Bunge) Kitagawa. Zhongguo Zhong Yao Za Zhi. 1993;18:362-383.
- 16. Levey AI. Immunological localization of M1-M5 muscarinic acetylcholine receptors in peripheral tissue and brain. Life Sci. 1993; 52: 441-448.
- Elgen RM, Hege SS, Watson N. Muscarinic receptor subtypes and smooth muscle function. Pharmacol Rev. 1996;48:531-565.
- Cohen M, Schenck K, Colbert W. Role of 5-HT2 receptors in serotonin-induced contraction of nonvascular smooth muscle. J Pharmacol Exp Ther. 1985;232:770-774.
- 19. Smith GJ, Lefebvre RA. Non adrenergic non cholinergic responses in the rat ileum. Eur J Pharmacol. 1996;303:79-87.

- 20. Ekblad E, Sandler F. Motor response in rat ileum evoked by nitric oxide donorsvs. Field stimulation: Modulation by pituitary adenylate cyclase inhibitors. J Pharmacol Exp Ther. 1997;283:23-28.
- 21. Sadraei H, Shokoohinia Y, Sajjadi SE, Mozafari M. Antispasmodic effects of *Prangos ferulacea* acetone extract and its main component osthole on ileum contraction. Res Pharm Sci. 2013;8:137-144.
- 22. Sadraei H, Asghari G, Emami S. Inhibitory effect of *Rosa damascena* Mill flower essential oil, geraniol and citronellol on rat ileum contraction. Res Pharm Sci. 2013;8:17-23.
- 23. Stefanska J, Sarniak A, Wlodarczyk A. Apocynin reduces reactive oxygen species concentrations in exhaled breath condensate in asthmatics. Exp Lung Res. 2012;38:90-99.
- 24. Impellizzeri D, Esposto E, Mazzon E. Effect of apocynin, a NADPH oxidase inhibitor, on acute lung inflammation. Biochem Pharmacol. 2011;81:636-648.
- 25. Impellizzeri D, Mazzon E, Esposito E. Effect of apocynin, an inhibitor of NADPH oxidase, in the inflammatory process induced by an experimental model of spinal cord injury. Free Radic Res. 2011;45:221-236.
- 26. Ahmad A, Mondello S, Di Paola R. Protective effect of apocynin, a NADPH-oxidase inhibitor, against contrastinduced nephropathy in the diabetic rats: a comparison with n-acetylcycteine. Eur J Pharmacol. 2012;674:397-406.